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(19) (CA) CANADIAN PATENT (12)

(54) Derivatized Oligonucleotides Having Improved Uptake and Other Properties

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(54) Title: DERIVATIZED OLIGONUCLEOTIDES HAVING IMPROVED UPTAKE AND OTHER PROPERTIES		
<p>(57) Abstract</p> <p>Linked nucleosides having at least one functionalized nucleotide that bears a substituent such as a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, a lipid soluble vitamin, an RNA cleaving complex, a metal chelator, a porphyrin, an alkylator, a pyrene, a hybrid photocleavage/intercalator, or an aryl azide photo-crosslinking agent exhibit increased cellular uptake and other properties. The substituent can be attached via a linking group or via a linking group. If at least a portion of the remaining linked nucleosides are 2'-deoxy-2'-fluoro, 2'-O-methoxy, 2'-O-ethoxy, 2'-O-propoxy, 2'-O-aminooxy or 2'-O-alkoxy nucleosides, the substituent can be attached via a linking group at any of the 3' or the 5' positions of the nucleoside or on the heterocyclic base of the nucleoside or on the inter-nucleotide linkage linking the nucleoside to an adjacent nucleotide.</p>		

2122030

1

DERIVATIZED OLIGONUCLEOTIDES HAVING IMPROVED UPTAKE  
AND OTHER PROPERTIES

CROSS REFERENCE TO RELATED APPLICATIONS

This application is related to Canadian patent application 2,073,500 filed January 11, 1991, Canadian patent application 2,089,377 filed on August 12, 1991 and Canadian patent application 2,140,428, filed on July 20, 1993.

FIELD OF THE INVENTION

10 This application is directed to sequence specific oligonucleotides that include functionalized nucleosides having substituents such as steroids, reporter molecules, reporter enzymes, non-aromatic lipophilic molecules, peptides, or proteins attached via linking groups.

BACKGROUND OF THE INVENTION

Messenger RNA (mRNA) directs protein synthesis. Antisense methodology is the complementary hybridization of relatively short oligonucleotides to mRNA or DNA such that the normal, essential functions of these intracellular nucleic acids are disrupted. Hybridization is the sequence-specific hydrogen bonding of oligonucleotides to RNA or single-stranded DNA via complementary Watson-Crick base pairs.



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2122030

2

The naturally occurring events that provide the disruption of the nucleic acid function, discussed by Cohen in *Oligonucleotides: Antisense Inhibitors of Gene Expression*, CRC Press, Inc., Boca Raton, FL (1989), are thought to be of 5 two types. The first, hybridization arrest, denotes a terminating event in which the oligonucleotide inhibitor binds to the target nucleic acid and thus prevents, by simple steric hindrance, the binding of essential proteins, most often ribosomes, to the nucleic acid. Methyl phosphonate 10 oligonucleotides (see, e.g., Miller, et al., *Anti-Cancer Drug Design* 1987, 2, 117) and  $\alpha$ -anomer oligonucleotides, the two most extensively studied antisense agents, are thought to disrupt nucleic acid function by hybridization arrest.

The second type of terminating event for antisense 15 oligonucleotides involves the enzymatic cleavage of targeted RNA by intracellular RNase H. A 2'-deoxyribofuranosyl oligonucleotide or oligonucleotide analog hybridizes with the targeted RNA to form a duplex that activates the RNase H enzyme to cleave the RNA strand, thus destroying the normal 20 function of the RNA. Phosphorothioate oligonucleotides provide the most prominent example of antisense agents that operate by this type of antisense terminating event.

Considerable research is being directed to the application of oligonucleotides and oligonucleotide analogs 25 as antisense agents for diagnostics, research reagents, and therapeutics. At least for therapeutic purposes, the antisense oligonucleotides and oligonucleotide analogs must be transported across cell membranes or taken up by cells to express activity. One method for increasing membrane or 30 cellular transport is by the attachment of a pendant lipophilic group.

Ramirez, et al., *J. Am. Chem. Soc.* 1982, 104:, 5483, introduced the phospholipid group 5'-O-(1,2-di-O-myristoyl-sn-glycero-3-phosphoryl) into the dimer TpT independently at the 35 3' and 5' positions. Subsequently Shea, et al., *Nuc. Acids Res.* 1990, 18, 3777, disclosed oligonucleotides having a 1,2-di-O-hexyldecyl-rac-glycerol group linked to a 5'-phosphate

2122030

3

on the 5'-terminus of the oligonucleotide. Certain of the Shea, et al authors disclosed these and other compounds in patent application WO 90/10448 published on September 20th, 1990. Another glucosyl phospholipid was disclosed by Guerra, et al., *Tetrahedron Letters* 1987, 28, 3581.

In other work, a cholesteryl group was attached to the inter-nucleotide linkage between the first and second nucleotides (from the 3' terminus) of an oligonucleotide.

This work is disclosed in United States Patent No. 4,958,013

10 and by Letsinger, et al., *Proc. Natl. Acad. Sci. USA* 1989, 86, 6553. The aromatic intercalating agent anthraquinone was attached to the 2' position of a sugar fragment of an oligonucleotide as reported by Yamana, et al., *Bioconjugate Chem.* 1990, 1, 319.

Lemairte, et al., *Proc. Natl. Acad. Sci. USA* 1986, 84, 648 and Leonetti, et al., *Bioconjugate Chem.* 1990, 1, 149, disclose modifying the 3' terminus of an oligonucleotide to include a 3'-terminal ribose sugar moiety. Poly(L-lysine) was linked to the oligonucleotide via periodate oxidation of this terminal ribose followed by reduction and coupling through a N-morpholine ring. Oligonucleotide-poly(L-lysine) conjugates are described in European published patent application 0,251,283 A 2 published on January 7, 1988 wherein the lysine residue was coupled to a 5' or 3' phosphate of the 5' or 3' terminal nucleotide of the oligonucleotide. A disulfide linkage has also been utilized at the 3' terminus of an oligonucleotide to link a peptide to the oligonucleotide, as

2122030

4

described by Corey, et al., *Science* 1987, 238, 1401; Zukermann, et al., *J. Am. Chem. Soc.* 1988, 110, 1614; and Corey, et al., *J. Am. Chem. Soc.* 1989, 111, 8524.

Nelson, et al., *Nuc. Acids Res.* 1989, 17, 7187 describe a linking reagent for attaching biotin to the 3'-terminus of an oligonucleotide. This reagent, N-Fmoc-O-DMT-3-amino-1,2-propanediol, is commercially available from Clontech Laboratories (Palo Alto, CA) under the name 3'-Amine on and from Glen Research Corporation (Sterling, VA) under the name 10 3'-Amino-Modifier\*. This reagent was also utilized to link a peptide to an oligonucleotide, as reported by Judy, et al., *Tetrahedron Letters* 1992, 32, 879. A similar commercial reagent (actually a series of linkers having various lengths of polymethylene connectors) for linking to the 5'-terminus of an oligonucleotide is 5'-Amino-Modifier\* C6, also from Glen Research Corporation. These compounds or similar ones were utilized by Krieg, et al., *Antisense Research and Development* 1991, 1, 161 to link fluorescein to the 5'-terminus of an oligonucleotide. Other compounds of interest have also been 20 linked to the 3'-terminus of an oligonucleotide. Asseline, et al., *Proc. Natl. Acad. Sci. USA* 1984, 81, 3297 described linking acridine on the 3'-terminal phosphate group of an poly (Tp) oligonucleotide via a polymethylene linkage. Haralambidis, et al., *Tetrahedron Letters* 1987, 28, 5199 reported building a peptide on a solid state support and then linking an oligonucleotide to that peptide via the 3' hydroxyl group of the 3' terminal nucleotide of the oligonucleotide.

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Chollet, *Nucleosides & Nucleotides* 1990, 9, 957 attached an Aminolink 2 (Applied Biosystems, Foster City, CA) to the 5' terminal phosphate of an oligonucleotide. They then used the bifunctional linking group SMPB (Pierce Chemical Co., Rockford, IL) to link an interleukin protein to the oligonucleotide.

An EDTA iron complex has been linked to the 5 position of a pyrimidine nucleoside as reported by Dreyer, et al., *Proc. Natl. Acad. Sci. USA* 1985, 82, 968. Fluorescein 10 has been linked to an oligonucleotide in the same manner, as reported by Haralambidis, et al., *Nucleic Acid Research* 1987, 15, 4857 and biotin in the same manner as described in PCT application WO 086/02929 published on May 22, 1986. Fluorescein, biotin and pyrene were also linked in the same manner as reported by Telser, et al., *J. Am. Chem. Soc.* 1989, 111, 6966. A commercial reagent, Amino-Modifier-dT\* from Glen Research Corporation, can be utilized to introduce pyrimidine nucleotides bearing similar linking groups into oligonucleotides.

Cholic acid linked to EDTA for use in  
radioscintigraphic imaging studies was reported by Betebenner,  
et al., *Bioconjugate Chem.* 1991, 2, 117; however, it is not  
known to link cholic acid to nucleosides, nucleotides or  
5 oligonucleotides.

#### OBJECTS OF THE INVENTION

It is an object of this invention to provide sequence-specific oligonucleotides having improved transfer across cellular membranes.

10 It is a further object of this invention to provide improvements in research and diagnostic methods and materials for assaying bodily states in animals, especially disease states.

15 It is an additional object of this invention to provide therapeutic and research materials having improved transfer and uptake properties for the treatment of diseases through modulation of the activity of DNA or RNA.

#### BRIEF DESCRIPTION OF THE INVENTION

20 In accordance with these and other objects evident from this specification, there are provided compounds that comprise a plurality of linked nucleosides wherein at least one of the nucleosides is functionalized at the 2'-position with a substituent such as, for example, a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, a lipid soluble vitamin, an RNA cleaving complex, a metal chelator, a porphyrin, an alkylator, a hybrid photonuclease/intercalator, a pyrene, or an aryl azide photo-crosslinking agent. Preferably, the substituent is connected to 2'-position using an intervening linking group.

In certain preferred embodiments of the invention, the substituents comprise a steroid molecule, biotin, a reporter enzyme or a fluorescein dye molecule. In these embodiments, 35 the steroid molecule is selected from the group consisting of cholic acid, deoxycholic acid, dehydrocholic acid, cortisone,

2122030

6

testosterone, cholesterol and digoxigenin with the most preferred steroid molecule being cholic acid. Preferred reporter enzymes include horseradish peroxidase and alkaline phosphatase.

5 In further preferred embodiments, the non-aromatic lipophilic molecule attached to the 2'-position comprises an alicyclic hydrocarbon, saturated or unsaturated fatty acid, wax, terpenoid, or polycyclic hydrocarbon, including adamantane and buckminsterfullerenes. Waxes according to the  
10 invention include monohydric alcohol esters of fatty acids and fatty diamides. Buckminsterfullerenes include soccer ball-shaped, cage molecules comprising varying numbers of covalently bound carbon atoms. Terpenoids include the C<sub>10</sub> terpenes, C<sub>20</sub> sesquiterpenes, C<sub>30</sub> diterpenes including vitamin A (retinol), retinic acid, retinal and dehydroretinol, C<sub>40</sub> triterpenes, C<sub>50</sub> tetraterpenes and other higher polyterpenoids.

In other preferred embodiments, peptides or proteins attached to the 2'-position comprise sequence-specific  
20 peptides and sequence-specific proteins, including phosphatases, peroxidases and nucleases.

Preferred linking molecules of the invention comprise N-aminoalkoxy linkers, N-aminoalkylamino linkers, heterobifunctional linkers or homobifunctional linkers. A particularly preferred linking molecule of the invention is a 5-aminopentoxy group.

In preferred embodiments of the invention at least a portion of the linked nucleosides are 2'-deoxy-2'-fluoro, 2'-methoxy, 2'-ethoxy, 2'-proxy, 2'-aminoalkoxy or 2'-allyloxy  
30 nucleosides. In other preferred embodiments of the invention the linked nucleosides are linked with phosphorothioate linking groups.

The invention also provides compounds that have a plurality of linked nucleosides. In preferred embodiments,  
35 at least one of the nucleosides is: (1) a 2'-functionalized nucleoside having cholic acid linked to its 2'-position; (2) a heterocyclic base functionalized nucleoside having cholic

acid linked to its heterocyclic base; (3) a 5' terminal nucleoside having cholic acid linked to its 5'-position; (4) a 3' terminal nucleoside having cholic acid linked to its 3'-position; or (5) an inter-strand nucleoside having cholic acid linked to an inter-strand linkage linking said inter-strand nucleoside to an adjacent nucleoside.

In certain embodiments of the invention having linked nucleosides, at least one linked nucleosides bears a 2'-deoxy/-2'-fluoro, 2'-O-C<sub>1</sub>-C<sub>20</sub>-alkyl, 2'-O-C<sub>2</sub>-C<sub>20</sub>-alkenyl, 2'-O-C<sub>2</sub>-C<sub>20</sub>-alkynyl, 2'-S-C<sub>1</sub>-C<sub>20</sub>-alkyl, 2'-S-C<sub>2</sub>-C<sub>20</sub>-alkenyl, 2'-S-C<sub>2</sub>-C<sub>20</sub>-alkynyl, 2'-NH-C<sub>1</sub>-C<sub>20</sub>-alkyl, 2'-NH-C<sub>2</sub>-C<sub>20</sub>-alkenyl, or 2'-NH-C<sub>2</sub>-C<sub>20</sub>-alkynyl substituent.

Further in accordance with the invention there is provided a method of increasing cellular uptake of a compound having a plurality of linked nucleosides that includes contacting an organism with a compound where the compound includes at least one nucleoside functionalized at the 2'-position with a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, and a lipid soluble vitamin. The compound can be included in a composition that further includes an inert carrier for the compound.

The invention also provides a method for enhancing the binding affinity and/or stability of an antisense 25 oligonucleotide comprising functionalizing the oligonucleotide with a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, and a lipid soluble vitamin.

2122030

8

## DETAILED DESCRIPTION OF THE INVENTION

Antisense therapeutics can be practiced in a variety of organisms ranging from unicellular prokaryotic and eukaryotic organisms to multicellular eukaryotic organisms. Any organism that utilizes DNA-RNA transcription or RNA-protein translation as a fundamental part of its hereditary, metabolic or cellular control is susceptible to antisense therapeutics and/or prophylactics. Seemingly diverse organisms such as bacteria, yeast, protozoa, algae, all plant and all higher animal forms, including warm-blooded animals, can be treated by antisense therapy. Further, since each of the cells of multicellular eukaryotes also includes both DNA-RNA transcription and RNA-protein translation as an integral part of its cellular activity, antisense therapeutics and/or diagnostics can also be practiced on such cellular populations. Furthermore, many of the organelles, e.g., mitochondria and chloroplasts, of eukaryotic cells also include transcription and translation mechanisms. As such, single cells, cellular populations or organelles can also be included within the definition of organisms that are capable of being treated with antisense therapeutics or diagnostics. As used herein, therapeutics is meant to include both the eradication of a disease state, killing of an organism, e.g., bacterial, protozoan or other infection, or control of erratic or harmful cellular growth or expression.

While we do not wish to be bound by any particular theory, it is believed that the presence of many nuclear proteins in the nucleus is due to their selective entry through the nuclear envelope rather than to their selective retention within the nucleus after entry. By this mechanism, the nucleus is able to selectively take up certain proteins and not others. The uptake is based upon the sequence of the peptide or protein, which provides a selective signal sequence that allows accumulation of the peptide or protein in the nucleus. One such peptide signal sequence is found as part of the SV40 large T-antigen. See, e.g., Dingwell, et al. *Ann. Rev. Cell Bio.* 1986, 2, 367; Yoneda, et al., *Experimental Cell*

Research 1987, 170, 439; and Wychowski, et al., J. Virol. 1986, 61, 3862.

According to the present invention a substituent such as a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, a lipid soluble vitamin, an RNA cleaving complex, a metal chelator, a porphyrin, an alkylator, a hybrid photonuclease/intercalator, or an aryl azide photocrosslinking agent is attached to at least one nucleoside in an antisense diagnostic or therapeutic agent to assist in the transfer of the antisense therapeutic or diagnostic agent across cellular membranes. Such antisense diagnostic or therapeutic agent is formed from a plurality of linked nucleosides of a sequence that is "antisense" to a region of an RNA or DNA that is of interest. Thus, one or more nucleoside of the linked nucleosides are "functionalized" to include a substituent linked to the nucleoside via linking group. For the purposes of identification, such functionalized nucleosides can be characterized as substituent-bearing (e.g., steroid-bearing) nucleosides. Linked nucleosides having at least one functionalized nucleoside within their sequence demonstrate enhanced antisense activity when compared to linked nucleoside that do not contain functionalized nucleoside. These "functionalized" linked nucleosides further demonstrate increased transfer across cellular membranes.

For the purposes of this invention, the terms "reporter molecule" and "reporter enzyme" include molecules or enzymes having physical or chemical properties that allow them to be identified in gels, fluids, whole cellular systems, broken cellular systems, and the like utilizing physical properties such as spectroscopy, radioactivity, colorimetric assays, fluorescence, and specific binding. Steroids include chemical compounds that contain a perhydro-1,2-cyclopantanophenanthrene ring system. Proteins and peptides are utilized in their usual sense as polymers of amino acids. Normally peptides are amino acid polymers that contain a fewer amino acid monomers

per unit molecule than proteins. Non-aromatic lipophilic molecules include fatty acids, esters, alcohols and other lipid molecules, as well as synthetic cage structures such as adamantane and buckminsterfullerenes that do not include 5 aromatic rings within their structure.

Particularly useful as steroid molecules are the bile acids, including cholic acid, deoxycholic acid and dehydrocholic acid. Other useful steroids are cortisone, digoxigenin, testosterone, cholesterol and cationic steroids 10 such as cortisone having a trimethylaminomethyl hydrazide group attached via a double bond at the 3 position of the cortisone rings. Particularly useful reporter molecules are biotin and fluorescein dyes. Particularly useful non-aromatic lipophilic molecules are alicyclic hydrocarbons, saturated and 15 unsaturated fatty acids, waxes, terpenes, and polycyclic hydrocarbons, including adamantane and buckminsterfullerenes. Particularly useful reporter enzymes are alkaline phosphatase and horseradish peroxidase. Particularly useful peptides and proteins are sequence-specific peptides and proteins, 20 including phosphodiesterase, peroxidase, phosphatase, and nuclease proteins. Such peptides and proteins include SV40 peptide, RNase A, RNase H and Staphylococcal nuclease. Particularly useful terpenoids are vitamin A, retinoic acid, retinal, and dehydroretinol.

25 Vitamins according to the invention generally can be classified as water soluble or lipid soluble. Water soluble vitamins include thiamine, riboflavin, nicotinic acid or niacin, the vitamin B<sub>6</sub> pyridoxal group, pantothenic acid, biotin, folic acid, the B<sub>12</sub> cobamide coenzymes, inositol, 30 choline and ascorbic acid. Lipid soluble vitamins include the vitamin A family, vitamin D, the vitamin E tocopherol family and vitamin K (and phytols). The vitamin A family, including retinoic acid and retinol, are absorbed and transported to target tissues through their interaction with specific 35 proteins such as cytosol retinol-binding protein type II (CRBP-II), Retinol-binding protein (RBP), and cellular retinol-binding protein (CRBP). These proteins, which have

been found in various parts of the human body, have molecular weights of approximately 15 kD. They have specific interactions with compounds of vitamin-A family, especially, retinoic acid and retinol.

5       The vitamin A family of compounds can be attached to oligonucleotides via acid or alcohol functionalities found in the various family members. For example, conjugation of an N-hydroxy succinimide ester of an acid moiety of retinoic acid to an amine function on a linker pendant to an oligonucleotide 10 resulted in linkage of vitamin A compound to the oligonucleotide via an amide bond. Also, retinol was converted to its phosphoramidite, which is useful for 5' conjugation.

$\alpha$ -Tocopherol (vitamin E) and the other tocopherols 15 (beta through zeta) can be conjugated to oligonucleotides to enhance uptake because of their lipophilic character. Also, the lipophilic vitamin, vitamin D, and its ergosterol precursors can be conjugated to oligonucleotides through their hydroxyl groups by first activating the hydroxyls groups to, 20 for example, hemisuccinate esters. Conjugation then is effected to an aminolinker pendant from the oligonucleotide. Other vitamins that can be conjugated to oligonucleotide aminolinkers through hydroxyl groups on the vitamins include thiamine, riboflavin, pyridoxine, pyridoxamine, pyridoxal, 25 deoxypyridoxine. Lipid soluble vitamin K's and related quinone-containing compounds can be conjugated via carbonyl groups on the quinone ring. The phytol moiety of vitamin K may also serve to enhance bind of the oligonucleotides to cells.

30       Pyridoxal (vitamin B<sub>6</sub>) has specific B<sub>6</sub>-binding proteins. The role of these proteins in pyridoxal transport has been studied by Zhang and McCormick, *Proc. Natl. Acad. Sci. USA*, 1991 **88**, 10407. Zhang and McCormick also have shown that a series of N-(4'-pyridoxyl)amines, in which several 35 synthetic amines were conjugated at the 4'-position of pyridoxal, are able to enter cells by a process facilitated by the B6 transporter. They also demonstrated the release of

these synthetic amines within the cell. Other pyridoxal family members include pyridoxine, pyridoxamine, pyridoxal phosphate, and pyridoxic acid. Pyridoxic acid, niacin, pantothenic acid, biotin, folic acid and ascorbic acid can be conjugated to oligonucleotides using N-hydroxysuccinimide esters that are reactive with aminolinkers located on the oligonucleotide, as described above for retinoic acid.

Other groups for modifying antisense properties include RNA cleaving complexes, pyrenes, metal chelators, porphyrins, alkylators, hybrid intercalator/ligands and photo-crosslinking agents. RNA cleavers include o-phenanthroline/Cu complexes and Ru(bipyridine),<sup>n</sup> complexes. The Ru(bpy)<sub>3</sub><sup>n+</sup> complexes interact with nucleic acids and cleave nucleic acids photochemically. Metal chelators are include EDTA, DTPA, and o-phenanthroline. Alkylators include compounds such as iodoacetamide. Porphyrins include porphine, its substituted forms, and metal complexes. Pyrenes include pyrene and other pyrene-based carboxylic acids that could be conjugated using the similar protocols.

Hybrid intercalator/ligands include the 20 photonuclease/intercalator ligand 6-[[[9-[(4-nitrobenzamido)hexyl]amino]acridin-4-yl]carbonyl]amino]hexanoylpentafluorophenyl ester. This compound has two noteworthy features: an acridine moiety that is an intercalator and a p-25 nitro benzamido group that is a photonuclease.

Photo-crosslinking agents include aryl azides such as, for example, N-hydroxysuccinimidyl-4-azidobenzoate (HSAB) and N-succinimidyl-6-(4'-azido-2'-nitrophenoxy-amino)hexanoate (SANPAN). Aryl azides conjugated to oligonucleotides effect 30 crosslinking with nucleic acids and proteins upon irradiation. They also crosslink with carrier proteins (such as K11 or BSA), raising antibody against the oligonucleotides.

A variety of linking groups can be used to connect the 35 substituents of the invention to nucleosides, nucleotides, and/or oligonucleotides. Certain linking groups, such as N-aminoalkoxy moieties and N-aminoalkylamine moieties, are particularly useful for linking steroid molecules or reporter

molecules to the 2'-position of a nucleoside. Many linking groups are commercially available, including heterobifunctional and homobifunctional linking moieties available from the Pierce Co. (Rockford, IL). 5 Heterobifunctional and homobifunctional linking moieties are particularly useful in conjunction with the N-aminoalkoxy and N-aminoalkylamino moieties to form extended linkers that connect peptides and proteins to nucleosides. Other commercially available linking groups are 5'-Amino-Modifier 10 C6 and 3'-Amino-Modifier reagents, both available from Glen Research Corporation (Sterling, VA). 5'-Amino-Modifier C6 is also available from ABI (Applied Biosystems Inc., Foster City, CA) as Aminolink-2, and 3'-Amino-Modifier is also available from Clontech Laboratories Inc. (Palo Alto, CA). A nucleotide 15 analog bearing a linking group pre-attached to the nucleoside is commercially available from Glen Research Corporation under the tradename "Amino-Modifier-dT." This nucleoside-linking group reagent, a uridine derivative having an [N(7-trifluoroacetylaminoheptyl)3-acrylamido] substituent group at the 5 20 position of the pyrimidine ring, is synthesized generally according to Jablonski, et al., *Nucleic Acid Research* 1986, 14, 6115. It is intended that the nucleoside analogs of the invention include adenine nucleosides functionalized with linkers on their N6 purine amino groups, guanine nucleosides 25 functionalized with linkers at their exocyclic N2 purine amino groups, and cytosine nucleosides functionalized with linkers on either their N4 pyrimidine amino groups or 5 pyrimidine positions.

Sequence-specific linked nucleosides of the invention 30 are assembled on a suitable DNA synthesizer utilizing either standard nucleotide precursors or nucleotide precursors that already bear linking moieties. Once synthesis of the sequence-specific linked nucleosides is complete, a substituent can be reacted with the linking moiety. Thus, the 35 invention preferably first builds a desired linked nucleoside sequence by known techniques on a DNA synthesizer. One or

2122030

14

more of the linked nucleosides are then functionalized or derivatized with a selected substituent.

Canadian patent applications 2,073,500; 2,089,377 and 2,140,428 disclose that incorporation of, for example, a 2'-O-methyl, 2'-O-ethyl, 2'-O-propyl, 2'-O-allyl, 2'-O-aminoalkyl or 2'-deoxy-2'-fluoro groups on the nucleosides of an oligonucleotide enhance the hybridization properties of the oligonucleotide. These applications also disclose that oligonucleotides containing phosphorothioate backbones have enhanced nuclease stability. The functionalized, linked nucleosides of the invention can be augmented to further include either or both a phosphorothioate backbone or a 2'-O-C<sub>1</sub>-C<sub>20</sub>-alkyl (e.g., 2'-O-methyl, 2'-O-ethyl, 2'-O-propyl), 2'-O-C<sub>2</sub>-C<sub>20</sub>-alkenyl (e.g., 2'-O-allyl), 2'-O-C<sub>2</sub>-C<sub>20</sub>-alkynyl, 2'-S-C<sub>1</sub>-C<sub>20</sub>-alkyl, 2'-S-C<sub>2</sub>-C<sub>20</sub>-alkenyl, 2'-S-C<sub>2</sub>-C<sub>20</sub>-alkynyl, 2'-NH-C<sub>1</sub>-C<sub>20</sub>-alkyl (2'-O-aminoalkyl), 2'-NH-C<sub>2</sub>-C<sub>20</sub>-alkenyl, 2'-NH-C<sub>2</sub>-C<sub>20</sub>-alkynyl or 2'-deoxy-2'-fluoro group.

An oligonucleotide possessing an amino group at its 5'-terminus is prepared using a DNA synthesizer and then is reacted with an active ester derivative of the substituent of the invention (e.g., cholic acid). Active ester derivatives are well known to those skilled in the art. Representative active esters include N-hydrosuccinimide esters, tetrafluorophenolic esters, pentafluorophenolic esters and pentachlorophenolic esters. For cholic acid, the reaction of

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the amino group and the active ester produces an oligonucleotide in which cholic acid is attached to the 5'-position through a linking group. The amino group at the 5'-terminus can be prepared conveniently utilizing the above-noted 5'-Amino-Modifier C6 reagent.

Cholic acid can be attached to a 3'-terminal amino group by reacting a 3'-amino modified controlled pore glass (sold by Clontech Laboratories Inc., Palo Alto, CA), with a cholic acid active ester.

10 Cholic acid can be attached to both ends of a linked nucleoside sequence by reacting a 3',5'-diamino sequence with the cholic acid active ester. The required oligonucleoside sequence is synthesized utilizing the 3'-Amino-Modifier\* and the 5'-Amino-Modifier\* C6 (or Aminolink-2\*) reagents noted above or by utilizing the above-noted 3'-amino modified controlled pore glass reagent in combination with the 5'-Amino-Modifier\* C2 (or Aminolink-2\*) reagents.

In even further embodiments of the invention, an oligonucleoside sequence bearing an aminolinker at the 2'-position of one or more selected nucleosides is prepared using a suitably functionalized nucleotide such as, for example, 5'-dimethoxytrityl-2'-O-( $\epsilon$ -phthalimidylaminopentyl)-2'-deoxyadenosine-3'-N,N-diisopropyl-cyanoethoxy phosphoramidite. See, e.g., Manoharan, et al., *Tetrahedron Letters*, 1991, 34, 7171 and above-referenced Canadian patent application 2,073,500 and 2,089,377. The nucleotide or nucleotides are attached to cholic acid or another substituent using an active

2122030

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ester or a thioisocyanate thereof. This approach allows the introduction of a large number of functional groups into a linked nucleoside sequence. Indeed each of the nucleosides can be so substituted.

In further functionalized nucleoside sequences of the invention, the heterocyclic base of one or more nucleosides is linked to a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, a lipid soluble vitamin, an RNA cleaving complex, a metal chelator, a porphyrin, an alkylator, a hybrid photonuclease/intercalator, or an aryl azide photo-crosslinking agent. Utilizing 5'-dimethoxytrityl-5-[N(7-trifluoroacetylaminohethyl)-3-acrylamido]-2'-deoxyuridine-3'-O-(methyl N,N-diisopropyl)-phosphoramide, as described by Jablonski, et al. above (also commercially available from Glen Research), the desired nucleoside, functionalized to incorporate a linking group on its heterocyclic base, is incorporated into the linked nucleoside sequence using a DNA synthesizer.

2122030 16

- Conjugation (linking) of reporter enzymes, peptides, and proteins to linked nucleosides is achieved by conjugation of the enzyme, peptide or protein to the above-described amino linking group on the nucleoside. This can be effected in several ways. A peptide- or protein- functionalized nucleoside of the invention can be prepared by conjugation of the peptide or protein to the nucleoside using EDC/sulfo-NHS (i.e., 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide/N-hydroxysulfosuccinimide) to conjugate the carboxyl end of the reporter enzyme, peptide, or protein with the amino function of the linking group on the nucleotide. Further, a linked nucleoside sequence of the invention can be prepared using EDC/sulfo-NHS to conjugate a carboxyl group of an aspartic or glutamic acid residue in the reporter enzyme, peptide or protein to the amino function of a linked nucleoside sequence.
- Preferably a reporter enzyme-, peptide-, protein-functionalized linked nucleoside sequence can be prepared by conjugation of the reporter enzyme, peptide or protein to the nucleoside sequence via a heterobifunctional linker such as m-maleimidobenzoyl-N-hydroxysulfosuccinimide ester (MBS) or succinimidyl 4-(N-maleimidomethyl)cyclohexane-1-carboxylate (SMCC) to link a thiol function on the reporter enzyme, peptide or protein to the amino function of the linking group on nucleoside sequence. By this mechanism, an oligonucleoside-maleimide conjugate is formed by reaction of the amino group of the linker on the linked nucleosides with the MBS or SMCC maleimide linker. The conjugate is then reacted with peptides or proteins having free sulphydryl groups.
- In a second preferred method, a reporter enzyme-, peptide-, protein-functionalized linked nucleoside sequence can be prepared by conjugation of the peptide or protein to the sequence using a homobifunctional linker such as disuccinimidyl suberate (DSS) to link an amino function on the peptide or protein to the amino group of a linker on the sequence. By this mechanism, an oligonucleoside-succinimidyl conjugate is formed by reaction of the amino group of the linker on the nucleoside sequence with a disuccinimidyl sub-

erate linker. The disuccinimidyl suberate linker couples with the amine linker on the sequence to extend the size of the linker. The extended linker is then reacted with amino groups such as, for example, the amine of lysine or other available 5 N-terminus amines, on reporter enzymes, peptides and proteins.

Additional objects, advantages, and novel features of this invention will become apparent to those skilled in the art upon examination of the following examples, which are not intended to be limiting.

10 For the following examples, anhydrous dimethylformamide, cholic acid and N-hydroxysuccinimide were purchased from Aldrich Chemical Co. (Milwaukee, WI), ethyl-3-(3-dimethylamino)propylcarbodiimide (EDAC or EDC) was obtained from JBL Scientific (San Luis Obispo, CA) as the free base 15 under the label EDAC or from Pierce (Rockford, IL) under the label EDC, Aminolink-2 was purchased from ABI and 3'-Amino-Modifier, 5'-Amino-Modifier C6 and Amino-Modifier dt reagents were purchased from Glen Research Corporation. NMR Spectra were run on a Varian Unity-400 instrument. Oligonucleotide 20 synthesis were performed on an Applied Biosystems 380 B or 394 DNA synthesizer following standard phosphoramidite protocols using reagents supplied by the manufacturer. When modified phosphoramidites were used, a longer coupling time (10-15 min) was employed. HPLC was performed on a Waters 600E instrument 25 equipped with a model 991 detector. Unless otherwise noted, for analytical chromatography the following conditions were employed: Hamilton PRP-1 column (15 x 2.5 cm); solvent A: 50mm TEAA, pH 7.0; solvent B: 45mm TEAA with 80% CH<sub>3</sub>CN; flow rate: 1.5ml/min; gradient: 5% B for the first 5 minutes, linear 30 (1%) increase in B every minute thereafter and for preparative purposes: Waters Delta Pak C-4 column; flow rate: 5ml/min; gradient: 5% B for the first 10 minutes, linear 1% increase for every minute thereafter.

All oligonucleotide sequences are listed in a standard 35 5' to 3' order from left to right.

**EXAMPLE 1****Cholic Acid N-Hydroxysuccinimide Ester (Compound 1)**

Anhydrous DMF (150ml) was added to a mixture of cholic acid (4.09g, 15mmol) and N-hydroxysuccinimide (5.25g, 45 mmol). The mixture was stirred in the presence of nitrogen. EDAC (4ml, 25mmol) was then added and this mixture was then stirred overnight. The solution was then evaporated to a gum and partitioned between 1:1 ethyl acetate and 4% NaHCO<sub>3</sub> solution (pH 7.9) (100ml each). The organic layer was washed 10 with saturated NaCl solution, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and evaporated to yield the title compound as a pale yellow foam (4.6g, 91%). <sup>13</sup>C NMR (DMSO-d<sub>6</sub>) δ 12.27, 16.71, 22.58, 22.80, 25.42, 26.19, 27.20, 28.49, 30.41, 30.56, 34.36, 34.82, 34.82, 35.31, 39.09, 39.09, 41.38, 41.53, 45.84, 46.05, 66.25, 70.45, 15 71.03, 169.28 and 170.16.

**EXAMPLE 2****Heterocyclic Base Cholic Acid End-Labeled Oligodeoxynucleotide**

An oligonucleotide of the sequence:

Oligomer 1: TTG CTT' .CCA TCT TCC TCG TC  
20 wherein T' represents a nucleotide functionalized to include a cholic acid linked via a linker to the heterocyclic base of a 2'-deoxyuridine nucleotide was prepared in a 1 μmol scale.  
Oligomer 1 is useful as an HPV antisense oligonucleotide.

**A. Preparation of Intermediate Linker**

25 The linker of the structure -CH=CH-C(O)-NH-(CH<sub>2</sub>)<sub>6</sub>-NH-C(O)-CF<sub>3</sub> was introduced via a suitable protected and activated 2'-deoxyuridine phosphoramidite intermediate obtained from Glen Research Corporation as Amino-Modifier-dT. The oligonucleotide bearing the linker thereon was deprotected and 30 purified by HPLC.

**B. Preparation of Cholic Acid Functionalized Oligonucleotide**

An aliquot of the linker bearing oligonucleotide of Example 2-A (approximately 100 O.D. units, 550 nmols) was 35 dissolved in 500 μl of 0.2M NaHCO<sub>3</sub> buffer and to this solution cholic acid N-hydroxysuccinimide ester (Compound 1, 75 mg, 149

19

( $\mu$ mol) was added and heated at 45°C overnight. It was then passed through a Sephadex G-25 (1.0 x 25 cm) column. Concentration of the oligonucleotide fractions to 2 ml followed by HPLC purification yielded the desired conjugate 5 wherein cholic acid is internally linked at C-5 of the heterocyclic base.

**EXAMPLE 3****5'-Terminus Cholic Acid End-Labeled Oligodeoxynucleotide**

A phosphorothioate oligonucleotide having cholic acid 10 attached to its 5'-terminus of the oligonucleotide sequence:

**Oligomer 2:**

5'-CHA-C<sub>2</sub>T,G<sub>3</sub>T,C<sub>4</sub>T,C<sub>5</sub>A<sub>6</sub>T,C<sub>7</sub>T,A<sub>8</sub>C<sub>9</sub>T<sub>10</sub> where CHA represents cholic acid and the subscript "g" represents a phosphorothioate inter-nucleotide backbone 15 linkage was prepared.

**A. Preparation of Intermediate Linker**

The oligonucleotide sequence having a 5'-terminus amino group was synthesized on a 3 x 1.0  $\mu$ mol scale in the standard manner on the DNA synthesizer utilizing phosphoramidite 20 methodology. The phosphorothioate intra-nucleotide backbone was formed using a phosphorothioate reagent (Beaucage reagent, i.e., 3H-1,2-benzodithioate-3-one 1,1-dioxide; see, Radhakrishnan, et al., J. Am. Chem. Soc. 1990, 112, 1253). The Aminolink-2 reagent was used at the last step of the 25 oligonucleotide synthesis. Deprotection with concentrated NH<sub>4</sub>OH for 16 hrs at 55°C yielded the 5'-aminolinker-oligonucleotide.

**B. Preparation of Cholic Acid Functionalized Oligonucleotide**

30 The crude 5'-aminolinker-oligonucleotide of Example 3-A (100 O.D. units, approximately 600 nmols based on the calculated extinction coefficient of  $1.6756 \times 10^5$  at 260 nm) was dissolved in freshly prepared NaHCO<sub>3</sub> buffer (500  $\mu$ l, 0.2M, pH 8.1) and treated with a solution of cholic acid N-35 hydroxysuccinimide ester (Compound 1, 75 mg, 149  $\mu$ mole) dissolved in 200  $\mu$ l of DMF. The reaction mixture was heated

2122030

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at 45°C overnight. It was then passed through a Sephadex G-25 (1.0 x 25 cm) column. Concentration of the oligonucleotide fractions to 2 ml followed by HPLC purification yielded 54 OD units of the desired conjugate (54% yield). HPLC retention times were: 37.42 for the unreacted oligonucleotide and any failure sequences produced during oligonucleotide synthesis and 54.20 for the final product.

**EXAMPLE 4****3'-Terminus Cholic Acid End Labeled Oligodeoxynucleotide**

10       A. **3'-Terminus Cholic Acid Oligonucleotide**  
A phosphorothioate oligonucleotide having cholic acid attached to its 3'-terminus of the oligonucleotide sequence:  
Oligomer 3:  
C<sub>1</sub>T<sub>2</sub>G<sub>3</sub> T<sub>4</sub>C<sub>5</sub>T<sub>6</sub> C<sub>7</sub>C<sub>8</sub>A<sub>9</sub> T<sub>10</sub>C<sub>11</sub>C<sub>12</sub> T<sub>13</sub>C<sub>14</sub>A<sub>15</sub> C<sub>16</sub>T<sub>17</sub> 3'-CHA  
15 wherein CHA represents cholic acid and the subscript "s" represents a phosphorothioate inter-nucleotide backbone linkage was prepared.

**1. Preparation of Intermediate Linker**

The oligonucleotide sequence having a 3'-terminus amino group was synthesized using 3'-amino modifier controlled pore glass (CPG) from Clontech Laboratories (Palo Alto, CA) as the solid support. The above-noted Beaucage reagent was utilized to form the phosphorothioate inter-nucleotide backbone. The synthesis was conducted in a "Trityl-Off" mode. The resultant solid support was deprotected with concentrated NH<sub>4</sub>OH for 16 hrs at 55°C. Purification on a Sephadex G-25 column yielded a 3'-amino functionalized phosphorothioate oligonucleotide of the specified oligonucleotide sequence.

**2. Preparation of Cholic Acid Functionalized**

30 **Oligonucleotide**  
The crude oligonucleotide of Example 4-A-1 (50 O.D. units, approximately 300 nmols) was reacted with cholic acid N-hydroxysuccinimide ester (Compound 1, 40 mg) as per the procedure of Example 3. HPLC retention times were 37.45 for 35 the starting oligonucleotide material and 51.68 for the product.

**B. 3'-Terminus Cholic Acid Oligonucleotide**

A phosphorothioate oligonucleotide having cholic acid attached to its 3'-terminus and of the oligonucleotide sequence:  
5           Oligomer 4:  
T<sub>3</sub>G<sub>4</sub>C<sub>5</sub> G<sub>6</sub>A<sub>7</sub>G<sub>8</sub> C<sub>9</sub>C<sub>10</sub>A<sub>11</sub> T<sub>12</sub>A<sub>13</sub>G<sub>14</sub> C<sub>15</sub>G<sub>16</sub>A<sub>17</sub> G<sub>18</sub>G<sub>19</sub>C<sub>20</sub> 3'-CHA  
wherein CHA represents cholic acid and the subscript "s" represents a phosphorothioate inter-nucleotide backbone linkage was prepared in the same manner as for the 10 oligonucleotide of Example 4-A-2.

**EXAMPLE 5**

3'-Terminus Cholic Acid & 5'-Terminus Cholic Acid Di-End-Labeled Oligodeoxynucleotide

A phosphorothioate oligonucleotide having cholic acid 15 attached to both of the 3'-terminus and the 5'-terminus of the oligonucleotide sequence:

5           Oligomer 5:  
5'-CHA C<sub>1</sub>T<sub>2</sub>G<sub>3</sub> T<sub>4</sub>C<sub>5</sub>T<sub>6</sub> C<sub>7</sub>C<sub>8</sub>A<sub>9</sub> T<sub>10</sub>C<sub>11</sub>T<sub>12</sub> T<sub>13</sub>C<sub>14</sub> G<sub>15</sub>T<sub>16</sub> 3'-CHA  
wherein CHA represents cholic acid and the subscript "s" 20 represents a phosphorothioate inter-nucleotide backbone linkage was synthesized on a 3 x 1.0 μmol scale. Oligomer 5 has the same sequence as Oligomer 1 except for the cholic acid functionalization.

**A. Preparation of Intermediate Linker**

25       The oligonucleotide synthesis was conducted using a 3'-amino modified controlled pore glass (CPG) from Clontech Laboratories as the solid support. Oligonucleotide synthesis was conducted utilizing phosphoramidite synthetic methodology and the Beaucage reagent to form a phosphorothioate inter-nucleotide backbone as per Example 4 above. Upon completion 30 of the basic sequence and while still on the DNA synthesizer, Aminolink-2 reagent was used to introduce a 5'-terminus amino functionality on to the oligonucleotide. Deprotection with concentrated ammonium hydroxide and purification on a Sephadex 35 G-25 column yielded 3',5'-diaminolinker oligonucleotide.

22

B. Preparation of Cholic Acid Functionalized Oligonucleotide

The crude di-aminolinker oligonucleotide (50 O.D. units, approximately 300 nmols, based on the calculated extinction coefficient of  $1.6756 \times 10^5$  at 260 nm) was dissolved in freshly prepared NaHCO<sub>3</sub> buffer (500 µl, 0.2M, pH 8.1) and treated with a solution of cholic acid N-hydroxysuccinimide ester (Compound 1, 50 mg, 98.6 µmols) dissolved in 200 µl of DMF. The reaction mixture was heated 10 at 45°C overnight. It was then passed through a Sephadex G-25 (1.0 x 25 cm) column. The oligonucleotide fractions were concentrated to 2 ml and purified by reverse phase HPLC. Retention times were 37.76 for unreacted oligonucleotide, 51.65 for 3'-cholic acid conjugated oligonucleotide, 54.34 for 15 5'-cholic acid conjugated oligonucleotide and 58.75 for 3',5'-di-cholic acid conjugated oligonucleotide. The 58.75 min. product was desalted on a Sephadex G-25 column to yield 11 O.D. units (22%) of the desired product.

EXAMPLE 6

20 3'-Terminus Cholic Acid or 5'-Terminus Cholic Acid Functionalized, 2'-O-Methyl Derivatized Oligodeoxynucleotides

Phosphorothioate oligonucleotides having cholic acid attached to either the 3'-terminus end or the 5'-terminus end of the oligonucleotide sequence and further being uniformly 25 functionalized to include a 2'-O-methyl group on each of the nucleotides of the oligonucleotide were synthesized. The following oligonucleotides having uniform 2'-O-methyl substitutions were synthesized:

30 Oligomer 6: 5'-CHA CCC AGG CUC AGA 3';  
Oligomer 7: 5' CCC AGG CUC AGA 3'-CHA; and  
Oligomer 8: 5'-CHA GAG CUC CCA GGC 3'.

A. Preparation of Intermediate Linker

Synthesis of the intermediate 5' or 3'-aminolinker oligonucleotide was conducted utilizing 2'-O-methyl phosphoramidite nucleotides available from Chemgenes Inc. (Needham, MA) and phosphoramidite chemistry as per Examples 3 and 4

23

above, respectively. Each of the intermediate oligonucleotides were deprotected in concentrated NH<sub>4</sub>OH, evaporated and de-salted on a Sephadex G-25 column.

B. Preparation of Cholic Acid Functionalized

5 Oligonucleotide

The resultant crude oligonucleotides from Example 6-A-1 (from 30 to 40 O.D. units, 250-350 nmols based on calculated extinction coefficients of  $1.1797 \times 10^5$ ,  $1.1777 \times 10^5$  and  $1.1481 \times 10^5$  at 260 nm, respectively for Oligomers 6, 7 and 8) 10 were dried and dissolved in 250  $\mu$ l of 0.2M NaHCO<sub>3</sub> buffer and treated with a solution of cholic acid N-hydroxysuccinimide ester (Compound 1, from 30 to 40 mg, 60 to 80  $\mu$ moles) dissolved in 500  $\mu$ l of 0.2 M NaHCO<sub>3</sub> buffer and 200  $\mu$ l DMF and heated between 40-45°C for 12-24 hrs. The reaction mixtures were 15 evaporated and dissolved in 2 ml of water and washed with 3 x 2 ml of ethyl acetate. The resultant aqueous solutions were purified by reverse phase HPLC. For Oligomer 6 the HPLC retention times were 34.65 for the starting oligonucleotide material and 58.75 for the product; for Oligomer 7 the HPLC 20 retention times were 37.23 for the starting oligonucleotide material and 55.32 for the product; and for Oligomer 8 the HPLC retention times were 34.99 for the starting oligonucleotide material and 56.98 for the product. The products were evaporated and desalted on a Sephadex G-25 25 column. The yield averaged about 20% in each case.

EXAMPLE 7

Oligonucleotides Having 2'-Protected-Amine Terminating Linking Group

A. Preparation of 5'-Dimethoxytrityl-2'-(O-Pentyl-N-30 phthalimido)-2'-Deoxyadenosine Phosphoramidite (Compound 2)

To introduce a functionalization at the 2' position of nucleotides within desired oligonucleotide sequences, 5'-Dimethoxytrityl-2'-(O-pentyl-N-phthalimido)-2'-deoxyadenosine phosphoramidite (Compound 2) was utilized to provide a linking 35 group attached to the 2' position of nucleotide components of an oligonucleotide. Compound 2 was synthesized as per the

2122030

24

procedures of Canadian patent application 2,073,550 identified above starting from adenosine. Briefly this procedure treats adenosine with NaN<sub>3</sub> in DMF followed by treatment with N-(5-bromopentyl)phthalimide. Further 5 treatment with (CH<sub>3</sub>)<sub>2</sub>SiCl, Ph-C(O)-Cl and NH<sub>4</sub>OH yields N6-benzyl protected 2'-pentyl-N-phthalimido functionalized adenosine. Treatment with DIPSA and CH<sub>2</sub>Cl<sub>2</sub> adds a DMT blocking group at the 5' position. Finally phosphorylation gives the desired phosphoramidite compound, Compound 2. Compound 2 was 10 utilized in the DNA synthesizer as a 0.09M solution in anhydrous CH<sub>3</sub>CN. Oligonucleotide synthesis was carried out in either an ABI 390B or 394 synthesizer employing the standard synthesis cycles with an extended coupling time of 10 minutes during coupling of Compound 2 into the oligonucleotide 15 sequence. Coupling efficiency of > 98% was observed for Compound 2 coupling.

B. 2'-Protected-Amine Linking Group Containing Phosphodiester Oligonucleotides

The following oligonucleotides having phosphodiester 20 inter-nucleotide linkages were synthesized:

Oligomer 9: 5' TAG 3';  
Oligomer 10: 5' CCA A G 3';  
Oligomer 11: 5' GGC TGA CTG CG 3';  
Oligomer 12: CTG TCT CCA TCC TCT TCA CT; and  
25 Oligomer 13: CTG TCT CCA TCC TCT TCA CT

wherein A' represents a nucleotide functionalized to incorporate a pentyl-N-phthalimido functionality. Oligomers 12 and 13 are antisense compounds to the E2 region of the bovine papilloma virus-1 (BPV-1). Oligomers 12 and 13 have 30 the same sequence as Oligomer 3 except for the 2' modification. The oligonucleotides were synthesized in either a 10 μmol scale or a 3 x 1 μmol scale in the "Trityl-On" mode. Standard deprotection conditions (30% NH<sub>4</sub>OH, 55°C, 24 hr) were employed. The oligonucleotides were purified by reverse phase 35 HPLC (Waters Delta-Pak<sup>\*</sup> C<sub>18</sub> 15 mm, 300A, 25x100 mm column equipped with a guard column of the same material). They were detritylated and further purified by size exclusion using a

\* Trade-mark

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Sephadex G-25 column. NMR analyses by both proton and phosphorus NMR confirmed the expected structure for the oligomers 9 and 10.

C. 2'-Protected-Amine Linking Group Containing  
5 Phosphorothioate Oligonucleotides

The following oligonucleotides having phosphorothioate inter-nucleotide linkages were synthesized:

Oligomer 14:

T<sub>1</sub>T<sub>2</sub>G<sub>1</sub> C<sub>2</sub>T<sub>3</sub>T<sub>4</sub> C<sub>3</sub>C<sub>4</sub>A<sup>\*</sup>, T<sub>1</sub>C<sub>2</sub>T<sub>3</sub> T<sub>2</sub>C<sub>3</sub>C<sub>4</sub> T<sub>3</sub>C<sub>5</sub>;

10 Oligomer 15:

T<sub>1</sub>G<sub>2</sub>G<sub>3</sub> G<sub>4</sub>A<sub>5</sub>G<sub>6</sub> C<sub>5</sub>C<sub>6</sub>A, T<sub>1</sub>A<sub>2</sub>G<sub>3</sub> C<sub>2</sub>G<sub>3</sub>A<sup>\*</sup>, G<sub>4</sub>G<sub>5</sub>C; and

Oligomer 16:

T<sub>1</sub>G<sub>2</sub>G<sub>3</sub> G<sub>4</sub>A<sup>\*</sup>G<sub>5</sub> C<sub>5</sub>C<sub>6</sub>A<sup>\*</sup>, T<sub>1</sub>A<sup>\*G<sub>2</sub> C<sub>2</sub>G<sub>3</sub>A<sup>\*</sup>, G<sub>4</sub>G<sub>5</sub>C</sup>

wherein A\* represents a nucleotide functionalized to incorporate a pentyl-N-phthalimido functionality and the subscript "\*" represents a phosphorothioate inter-nucleotide backbone linkage. Oligomer 14 is an antisense compound directed to the E2 region of the bovine papilloma virus-1 (BPV-1). Oligomers 15 and 16 are antisense compounds to ICAM. Oligomer 14 has the same sequence as Oligomer 3 except for the 2' modification whereas Oligomers 15 and 16 have the same sequence as Oligomer 4 except for the 2' modification. These oligonucleotides were synthesized as per the method of Example 7-B except during the synthesis, for oxidation of the 25 phosphate moieties, the Beaucage reagent (see Example 3 above) was used as a 0.24 M solution in anhydrous CH<sub>3</sub>CN solvent. The oligonucleotides were synthesized in the "Trityl-On" mode and purified by reverse phase HPLC utilizing the purification procedure of Example 7-B.

30 D. 2'-O-Methyl Derivatized, 2'-Protected-Amine Linking  
Group Containing RNA Oligonucleotides

The following oligonucleotides having 2'-O-methyl groups on each nucleotide not functionalized with a 2'-protected amine functionalization were synthesized:

35 Oligomer 17: CCA A\*GC CUC AGA; and  
Oligomer 18: CCA CGG UCA GA\*

wherein A\* represents a nucleotide functionalized to incorporate a pentyl-N-phthalimido functionality and where the remaining nucleotides except the 3'-terminus nucleotide are each 2'-O-methyl derivatized nucleotides. The 3'-terminus nucleotide in both Oligomers 17 and 18 is a 2'-deoxy nucleotide. Both Oligomers 17 and 18 are antisense compounds to the HIV-1 TAR region. The oligonucleotides were synthesized as per the method of Example 6 utilizing compound 2 for introduction of the nucleotides containing the pentyl-N-phthalimido functionality and appropriate 2-O-methyl phosphoramidite nucleotides from Chemgenes Inc. (Needham, MA) for the remaining RNA nucleotides. The 3'-terminus terminal 2'-deoxy nucleotides were standard phosphoamidites utilized for the DNA synthesizer. The oligonucleotides were deprotected and purified as per the method of Example 7-B.

**EXAMPLE 8**

**Functionalization of Oligonucleotides At the 2' Position**

**A. Functionalization with Biotin**

**1. Single Site Modification**

About 10 O.D. units ( $A_{260}$ ) of Oligomer 12 (see Example 7) (approximately 60 nmols based on the calculated extinction coefficient of  $1.6756 \times 10^5$ ) was dried in a microfuge tube. The oligonucleotide was dissolved in 200  $\mu$ l of 0.2 M NaHCO<sub>3</sub> buffer and D-biotin-N-hydroxysuccinimide ester (2.5 mg, 7.3 nmols) (Sigma, St. Louis, MO) was added followed by 40  $\mu$ l DMF. The solution was let stand overnight. The solution was applied to a Sephadex G-25 column (0.7 x 15 cm) and the oligonucleotide fractions were combined. Analytical HPLC showed nearly 85% conversion to the product. The product was purified by HPLC (Waters 600E with 991 detector, Hamilton PRP-1 column 0.7 x 15 cm; solvent A: 50 mM TEAA pH 7.0; B: 45 mM TEAA with 80% acetonitrile: 1.5 ml flow rate: Gradient: 5% B for first 5 mins., linear (1%) increase in B every minute thereafter) and further desalting on Sephadex G-25 to give the oligonucleotide:

Oligomer 19: CTG TCT CCA' TCC TCT TCA CT

2122030  
27

wherein A' represents a nucleotide functionalized to incorporate a biotin functionality linked via a 2'-O-pentyl-amino linking group to the 2' position of the designated nucleotide. HPLC retention times are shown in Table 1 below.

5           2. Multiple Site Modification

About 10 O.D. units ( $A_{260}$ ) of Oligomer 13 (see Example 7, approximately 60 nmols) was treated utilizing the method of Example 8-A-1 with D-biotin-N-hydroxysuccinimide ester (5 mg) in 300  $\mu$ l of 0.2 M NaHCO<sub>3</sub> buffer/ 50  $\mu$ l DMF. Analytical HPLC showed 65% of double labeled oligonucleotide product and 30% of single labeled products (from the two available reactive sites). HPLC and Sephadex G-25 purification gave the oligonucleotide:

Oligomer 20: CTG TCT CCA' TCC TCT TCA' CT  
15       wherein A' represents nucleotides functionalized to incorporate a biotin functionality linked via a 2'-O-pentyl-amino linking group to the 2' position of the designated nucleotide. HPLC retention times for this product (and its accompanying singly labeled products) are shown in Table 1  
20       below.

B. Functionalization with Fluorescein

1. Single Site Modification

A 1M Na<sub>2</sub>CO<sub>3</sub>/1M NaHCO<sub>3</sub> buffer (pH 9.0) was prepared by adding 1M NaHCO<sub>3</sub> to 1M Na<sub>2</sub>CO<sub>3</sub>. 200  $\mu$ l of this buffer was added to 10 O.D. units of Oligomer 12 (see Example 7) in a microfuge tube. 10 mg of fluorescein-isocyanate in 500  $\mu$ l DMF was added to give a 0.05 M solution. 100  $\mu$ l of the fluorescein solution was added to the oligonucleotide solution in the microfuge tube. The tube was covered with aluminum foil and let stand overnight. The reaction mixture was applied to a Sephadex G-25 column (0.7 x 20 cm) that had been equilibrated with 25% (v/v) ethyl alcohol in water. The column was eluted with the same solvent. Product migration could be seen as a yellow band well separated from dark yellow 30 band of the excess fluorescein reagent. The fractions showing absorption at 260 nm and 485 nm were combined and purified by HPLC as per the purification procedure of Example 8-A-1.  
35

Analytical HPLC indicated 61% of the desired doubly functionalized oligonucleotide. The product was lyophilized and desalted on Sephadex to give the oligonucleotide:

Oligomer 21: CTG TCT CCA' TCC TCT TCA CT

5 wherein A' represents a nucleotide functionalized to incorporate a fluorescein functionality linked via a 2'-O-pentyl-amino linking group to the 2' position of the designated nucleotide. HPLC retention times are shown in Table 1 below.

10 2. Multiple Site Modification

10 O.D. units ( $A_{260}$ ) of Oligomer 13 (from Example 7) was dissolved in 300  $\mu$ l of the 1M  $Na_2HCO_3$ / 1M  $Na_2CO_3$  buffer of Example 8-B-1 and 200  $\mu$ l of the fluorescein-isothiocyanate stock solution of Example 8-B-1 was added. The resulting 15 solution was treated as per Example 8-B-1. Analytical HPLC indicated 61% of doubly labeled product and 38% of singly labeled products. Work up of the reaction gave the oligonucleotide:

Oligomer 22: CTG TCT CCA' TCC TCT TCA' CT

20 wherein A' represents nucleotides functionalized to incorporate a fluorescein functionality linked via a 2'-O-pentyl-amino linking group to the 2' position of the designated nucleotide. HPLC retention times are shown in Table 1 below.

25 C. Functionalization with Cholic Acid

1. Single Site Modification

10 O.D. units ( $A_{260}$ ) of Oligomer 12 (see Example 7) was treated with cholic acid-NHS ester (compound 1, 5 mg, 9.9  $\mu$ moles) in 200  $\mu$ l of 0.2 M  $NaHCO_3$  buffer/ 40  $\mu$ l DMF. The 30 reaction mixture was heated for 16 hrs at 45°C. The product was isolated as per the method of Example 8-A-1. Analytical HPLC indicated > 85% product formation. Work up of the reaction gave the oligonucleotide:

Oligomer 23: CTG TCT CCA' TCC TCT TCA CT

35 wherein A' represents a nucleotide functionalized to incorporate a cholic acid functionality linked via a 2'-O-pentyl-amino linking group to the 2' position of the

designated nucleotide. HPLC retention times are shown in Table 1 below.

2. Multiple Site Modification

10 O.D. units ( $A_{260}$ ) of Oligomer 13 (see Example 7) was 5 treated with cholic acid-NHS ester (Compound 1, 10 mg, 19.8  $\mu$ moles) in 300  $\mu$ l of 0.2 M NaHCO<sub>3</sub> buffer/ 50  $\mu$ l DMF. The reaction mixture was heated for 16 hrs at 45°C. The product was isolated as per the method of Example 8-A-1. Analytical HPLC revealed 58% doubly labeled product, 17% of a first 10 singly labeled product and 24% of a second singly labeled product. Work up as per Example 8-A-1 gave the oligonucleotide:

Oligomer 24: CTG TCT CCA' TCC TCT TCA' CT  
wherein A' represents nucleotides functionalized to 15 incorporate a cholic acid functionality linked via a 2'-O-pentyl-amino linking group to the 2' position of the designated nucleotide. HPLC retention times are shown in Table 1 below.

D. Functionalization with Digoxigenin

20 1. Single Site Modification  
10 O.D. units ( $A_{260}$ ) of Oligomer 12 (see Example 7) was treated with digoxigenin-3-O-methylcarbonyl- $\epsilon$ -aminocaproic N-hydroxy succinimide ester (Boehringer Mannheim Corporation, Indianapolis, IN) in 200  $\mu$ l of 0.1 M borate pH 8.3 buffer/ 40 25  $\mu$ l DMF. The reaction mixture was let stand overnight. The product was isolated as per the method of Example 8-A-1. Work up of the reaction gave the oligonucleotide:

Oligomer 25: CTG TCT CCA' TCC TCT TCA CT  
wherein A' represents a nucleotide functionalized to 30 incorporate a digoxigenin functionality linked via a 2'-O-pentyl-amino linking group to the 2' position of the designated nucleotide. HPLC retention times are shown in Table 1 below.

2. Multiple Site Modification

35 10 O.D. units ( $A_{260}$ ) of Oligomer 13 (see Example 7) was treated with digoxigenin-3-O-methylcarbonyl- $\epsilon$ -aminocaproic N-hydroxy succinimide ester (Boehringer Mannheim Corporation,

2122030

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Indianapolis, IN) in 300  $\mu$ l of 0.1 M borate pH 8.3 buffer / 50  $\mu$ l DMF. The reaction mixture was let stand overnight. The product was isolated as per the method of Example 8-A-1. Work up as per Example 8-A-1 gave the oligonucleotide:

5           Oligomer 26: CTG TCT CCA<sup>A</sup> TCC TCT TCA<sup>A</sup> CT  
 wherein A<sup>A</sup> represents nucleotides functionalized to incorporate a cholic acid functionality linked via a 2'-O-pentyl-amino linking group to the 2' position of the designated nucleotide. HPLC retention times are shown in  
 10 Table 1 below.

TABLE I  
 HPLC Retention Times Of Oligonucleotides Functionalized  
 At 2' Position

15	Oligomer	Retention Time Minutes	
		Mono Substitution	Multiple Substitution
	Oligomer 12 <sup>1</sup>	21.78	
	Oligomer 13 <sup>1</sup>		22.50
	Oligomer 19 <sup>2</sup>	23.58	
20	Oligomer 20 <sup>2</sup>		24.16 <sup>b</sup> 25.19 <sup>b</sup>
	Oligomer 21 <sup>3</sup>	26.65	26.99 <sup>b</sup> 29.33 <sup>b</sup>
	Oligomer 22 <sup>3</sup>		27.55 <sup>b</sup>
	Oligomer 23 <sup>4</sup>	30.10	
25	Oligomer 24 <sup>4</sup>		30.38 <sup>b</sup> 37.00 <sup>b</sup>
	Oligomer 25 <sup>5</sup>	28.06	32.22 <sup>b</sup>
	Oligomer 26		28.14 <sup>a</sup> 33.32 <sup>b</sup> 29.24 <sup>a</sup>

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Conditions: Waters 600E with 991 detector, Hamilton PRP-1 column, 0.7 x 15 cm; solvent A: 50 mM TEAA pH 7.0; B: 45 mM TEAA with 80% acetonitrile: 1.5 ml flow rate: Gradient: 5% B for first 5 mins., linear (1%) increase in B every minute thereafter;

<sup>a</sup> Mono conjugated minor product;

<sup>b</sup> Doubly conjugated major product;

<sup>1</sup> Parent Oligonucleotide - no 2' functionalization;

<sup>2</sup> 2' Biotin functionalization;

<sup>3</sup> 2' Fluorescein functionalization;

<sup>4</sup> 2' Cholic Acid functionalization; and

<sup>5</sup> 2' Digoxigenin functionalization.

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**EXAMPLE 9****Functionalisation Of Oligonucleotide At the 2' Position with Reporter Enzymes, Peptides and Proteins****A. Use of Heterobifunctional Linker****5 1. Synthesis of Oligonucleotide-Maleimide Conjugate**

Oligomer 12 (Example 7) (100 O.D. units, 600 nmols) is lyophilized in a 5 ml pear-shaped flask. Sulfo-SMCC reagent, Pierce Chemical Co. (Rockford, IL) (16 mg, 46  $\mu$ moles) is dissolved in phosphate buffer (800  $\mu$ l, 0.1M, pH 7.0) and added to the oligonucleotide bearing flask. An additional 200  $\mu$ l of buffer are used to wash the reagent and transfer it to the oligonucleotide flask. The contents of the flask are stirred overnight and loaded on to a Sephadex G-25 column (1 x 40 cm) equipped with a fraction collector. The oligonucleotide-maleimide conjugate containing fractions are collected and tested by analytical HPLC for separation from other NHS type products.

**2. Synthesis of Oligonucleotide-Peptide Conjugate**

20 An aliquot of the oligonucleotide-maleimide conjugate of Example 9-A-1 (about 50 O.D. units, 300 nmols) is lyophilized in a microtube tube. SV40 peptide (pro-asp-lys-lys-arg-lys-cys) (2.5 mg, about 2.5  $\mu$ moles) is taken up in phosphate buffer (800  $\mu$ l, 0.1 M, pH 7.0) and added to the 25 oligonucleotide-maleimide conjugate containing tube. The contents of the tube are stirred overnight under an argon atmosphere. The reaction mixture is passed through a Sephadex G-25 column and the oligonucleotide-peptide conjugate fractions are identified by HPLC. Isolation of the product 30 from product-bearing fractions via HPLC and desalting on Sephadex G-25 will yield an oligonucleotide of the sequence:

Oligomer 27: CTC TCT CCA<sup>A</sup> TCC TCT TCA CT

wherein A represents a nucleotide functionalized to incorporate a SV40 peptide functionality linked via a 2'-O-35 pentyl-amino-sulfo-SMCC (sulfosuccinimidyl-4-(N-maleimidomethyl)cyclohexane-1-carboxylate) linking group to the 2' position of the designated nucleotide.

2122030

32

## B. Use of Homobifunctional Linker

## 1. Synthesis of Oligonucleotide-Disuccinimidyl

## Suberate Conjugate

An aliquot (10 O.D. units, 60 nmols) of Oligomer 12 (Example 7) is evaporated to dryness and is dissolved in freshly prepared 0.1 M NaHCO<sub>3</sub>/50 mM EDTA (100 µl, pH 8.25). The solution is then treated with a solution of DSS, Pierce Chemical Co. (Rockford, IL) (2.6 mg, 7 µmol) in 200 µl DMSO. The solution is stored at room temperature for 15 minutes and 10 then immediately applied to a Sephadex G-25 column (1 x 40 cm) that is previously packed and washed with water at 4°C. The oligonucleotide fractions are combined immediately in a 25 ml pear-shaped flask and are rapidly frozen in dry ice/isopropyl alcohol and lyophilized to a powder.

## 15 2. Synthesis of Oligonucleotide-Protein Conjugate

A solution of calf intestinal alkaline phosphatase (Boehringer Mannheim) (20.6 mg, 2.06 ml, 147 nmol) is spun at 4°C in a Centricon microconcentrator at 6000 rpm until the volume is less than 50 µl. It is then redissolved in 1 ml of 20 cold Tris buffer (pH 8.5, 0.1M containing 0.1 NaCl and 0.05 M MgCl<sub>2</sub>) and concentrated twice more. Finally the concentrate is dissolved in 400 µl of the same buffer. This solution is added to the activated oligonucleotide from Example 9-B-1 and the solution stored for 18 hrs at room temp. The product is 25 diluted to approximately 30 ml and applied to a Sephadex G-25 column (1 x 20 cm, chloride form) maintained at 4°C. The column is eluted with 50 mM Tris-Cl pH 8.5 until the UV absorbance of the fractions eluted reach near zero values. The column is then eluted with a NaCl salt gradient 0.05 M to 30 0.75 M (150 ml each). The different peaks are assayed for both oligonucleotide and alkaline phosphatase activity and the product bearing fractions are combined. Typically the first peak will be excess enzyme, the second peak the oligonucleotide-protein conjugate and the third peak unreacted 35 oligonucleotide. Isolation of the product from the product-bearing fractions via HPLC and desalting on Sephadex G-25 will yield an oligonucleotide of the sequence:

Oligomer 28: CTG TCT CCA TCC TCT TCA CT  
wherein A' represents a nucleotide functionalized to incorporate an alkaline phosphatase functionality linked via an 2'-O-pentyl-amino-sulfo-SMCC (sulfosuccinimidyl-4-(N-maleimidomethyl)cyclohexane-1-carboxylate) linking group to the 2' position of the designated nucleotide.

EXAMPLE 10

Heterocyclic Base Peptide-Conjugated Oligonucleotides

Utilizing the method of Example 9-A-1 the intermediate 10 amino linker oligonucleotide of Example 2-A is reacted with sulfo-SMCC reagent. The isolated oligonucleotide-maleimide conjugate is then further reacted with SV40 peptide as per Example 9-A-2. This will give an oligonucleotide of the structure:

15 Oligomer 29: TTG CTC' CCA TCT TCC TCG TC  
wherein T' represents a nucleotide functionalized to include a peptide linked via an extended linker to the heterocyclic base of a 2'-deoxyuridine nucleotide.

EXAMPLE 11

3'-Terminus Protein-Conjugated Oligonucleotides

Utilizing the method of Example 9-B-1 the 2'-O-methyl derivatized intermediate amino linker oligonucleotide of Example 6-A is reacted with DSS reagent. The isolated oligonucleotide-disuccinimidyl suberate conjugate is then 25 further reacted with a lysine containing Nuclease RNase H using the method of Example 9-B-2. This will give an oligonucleotide of the structure:

Oligomer 30: C<sub>1</sub>C<sub>2</sub>C<sub>3</sub>A<sub>1</sub>G<sub>2</sub>G<sub>3</sub>C<sub>4</sub>U<sub>5</sub>C<sub>6</sub>A<sub>7</sub>G<sub>8</sub>A-3'-protein  
wherein protein represents RNase H, the subscript "s" 30 represents a phosphorothioate inter-nucleotide backbone linkage and each of the nucleotides of the oligonucleotide includes a 2'-O-methyl group thereon.

2122030

34

**EXAMPLE 12****5'-Terminus Protein-Conjugated 2'-O-Methyl Derivatized Oligonucleotides**

Utilizing the method of Example 9-B-1 the 2'-O-methyl derivatized intermediate amino linker oligonucleotide of Example 6-A (Oligomer 6) is reacted with DSS reagent. The isolated oligonucleotide-disuccinimidyl suberate conjugate is then further reacted with a lysine containing Staphylococcal Nuclease using the method of Example 9-B-2. This will give an oligonucleotide of the structure:

Oligomer 31: 5'-protein-C<sub>s</sub>C<sub>s</sub>C<sub>s</sub>A<sub>s</sub>G<sub>s</sub>G<sub>s</sub>C<sub>s</sub>U<sub>s</sub>C<sub>s</sub>A<sub>s</sub>G<sub>s</sub>A 3'  
wherein protein represents Staphylococcal Nuclease, the subscript "s" represents a phosphorothioate inter-nucleotide backbone linkage and each of the nucleotides of the oligonucleotide includes a 2'-O-methyl group thereon.

**PROCEDURE A****Confirmation of Structure of Functionalized Oligonucleotides Containing A Tethered 2'-Amino Moiety**

Oligonucleotides of the invention were digested with snake venom phosphodiesterase and calf-intestine alkaline phosphatase to their individual nucleosides. After digestion, the nucleoside composition was analyzed by HPLC. The HPLC analysis established that functionalized nucleotide compounds having the tethered 2'-amino moiety thereon were correctly incorporated into the oligonucleotide.

Snake venom phosphodiesterase [Boehringer-Mannheim cat. #108260, 1 mg (1.5 units)/0.5 ml] and alkaline phosphatase from calf intestine (1 unit/microliter, Boehringer-Mannheim cat. # 713023) in Tris-HCl buffer (pH 7.2, 50 mM) were used to digest the oligonucleotides to their component nucleosides. To 0.5 O.D. units of oligonucleotide in 50  $\mu$ l buffer (nearly 40  $\mu$ M final concentration for a 20 mer) was added 5  $\mu$ l of snake venom phosphodiesterase (nearly 0.3 units/mL, final concentration) and 10  $\mu$ l of alkaline phosphatase (app. 150 units/mL, final concentration). The reaction mixture was incubated at 37°C for 3 hours. Following incubation, the reaction mixture was analyzed by HPLC using a reverse phase

2122030  
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analytical column (app. 30 x 2.5 cm); solvent A: 50 mM TEAA pH 7; solvent B: acetonitrile; gradient 100% for 10 mins, then 5% B for 15 mins, then 10% B and then wash. The results of these digestion are shown in Table 2 for representative 5 oligonucleotides.

TABLE 2  
OLIGONUCLEOTIDE ANALYSIS VIA ENZYMATIC DIGESTION

	Oligomer	Abs. max.	Observed Ratios**				
			267	252	267	260	A
10	Oligomer 10		2	1	1		
	Oligomer 11		3	5	2	1	
15	Oligomer 12		9	1	8	1	1
	Oligomer 13		9	1	8	2	

\* Nucleoside having 2'-O-linker attached thereto; and  
\*\* Corrected to whole numbers.

As is evident from Table 2, the correct nucleoside ratios are observed for the component nucleotides of the test oligonucleotides.

#### PROCEDURE B

#### Determination of Melting Temperatures ( $T_m$ 's) of Cholic Acid Oligonucleotide Conjugates

The relative ability of oligonucleotides to bind to their complementary strand is compared by determining the melting temperature of the hybridization complex of the oligonucleotide and its complementary strand. The melting temperature ( $T_m$ ), a characteristic physical property of double helices, denotes the temperature in degrees centigrade at which 50% helical versus coil (un-hybridized) forms are present. This is measured by using the UV spectrum to determine the formation and breakdown (melting) of hybridization. Base stacking, which occurs during hybridization, is accompanied by a reduction in UV absorption (hypochromicity). Consequently a reduction in UV absorption indicates a higher  $T_m$ . The higher the  $T_m$ , the greater the strength of the binding of the strands. Non-Watson-Crick base pairing has a strong destabilizing effect on the  $T_m$ . Consequently, absolute

fidelity of base pairing is necessary to have optimal binding of an antisense oligonucleotide to its targeted RNA.

1. Terminal End Conjugates

a. Synthesis

5 A series of oligonucleotides were synthesized utilizing standard synthetic procedures (for un-functionalized oligonucleotides) or the procedure of Example 3-A above for oligonucleotides having a 5'-terminus amino linker bearing oligonucleotide or the procedure of Example 3-B for 5'-terminus cholic acid-bearing oligonucleotides. Each of the oligonucleotides had the following 5'-LO antisense sequence: 10 5' TCC AGG TGT CCC CAT C 3'. The nucleotides were synthesized on a 1.0  $\mu$ mol scale. Oligomer 32 was the parent compound having normal phosphodiester inter-nucleotide linkages. 15 Oligomer 33 incorporated phosphorothioate inter-nucleotide linkages in the basic oligonucleotide sequence. Oligomer 34 is an intermediate oligonucleotide having a 5'-aminolink at the 5'-terminus of the basic oligonucleotide sequence and Oligomer 35 was a similar 5'-aminolink compound incorporating 20 phosphorothioate inter-nucleotide linkages. Oligomer 36 is a 5'-terminus cholic acid conjugate of the basic phosphodiester oligonucleotide sequence while Oligomer 37 is a similar 5'-cholic acid conjugate incorporating phosphorothioate inter-nucleotide linkages. Oligomers 32 and 25 33 were synthesized in a "Tritly-On" mode and were purified by HPLC. Oligomers 34 and 35 were synthesized as per Example 30-A above without or with Beaucage reagent treatment, to yield phosphodiester or phosphorothioate inter-nucleotide linkages, respectively. Oligomers 36 and 37 were prepared 30 from samples of Oligomers 34 and 35, respectively, utilizing a solution of cholic acid N-hydroxysuccinimide ester (Compound 1) 1 dissolved in DMF as per Example 3-B. Oligomers 36 and 37 were purified by HPLC. The products were concentrated and desalted in a Sephadex G-25 column. Gel electrophoresis 35 analyses also confirmed a pure product with the pure conjugate moving slower than the parent oligonucleotide or 5'-amino functionalized oligonucleotide.

2122030  
37

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**b. Melting Analysis**

The test oligonucleotides [either the phosphodiester, phosphorothioate, cholic acid conjugated phosphodiester, cholic acid conjugated phosphorothioate or 5'-aminolink 5 intermediate phosphodiester or phosphorothioate oligonucleotides of the invention or otherwise] and either the complementary DNA or RNA oligonucleotides were incubated at a standard concentration of 4  $\mu$ M for each oligonucleotide in buffer (100 mM NaCl, 10 mM Na-phosphate, pH 7.0, 0.1 mM EDTA).  
 10 Samples were heated to 90 degrees C and the initial absorbance taken using a Guilford Response II spectrophotometer (Corning). Samples were then slowly cooled to 15 degrees C and then the change in absorbance at 260 nm was monitored during the heat denaturation procedure. The temperature was  
 15 elevated 1 degree/absorbance reading and the denaturation profile analyzed by taking the 1st derivative of the melting curve. Data was also analyzed using a two-state linear regression analysis to determine the Tm's. The results of these tests are shown in Table 3 as are the HPLC retention  
 20 times of certain of the test compounds.

TABLE 3

Melting Temperature Of The Hybridization Complex Of The Oligonucleotide And Its Complementary Strand

	25 Oligomer	Tm**		HPLC Ret. Time* minutes
		DNA	RNA	
30	32	62.6	62.0	-----
	33	55.4	54.9	-----
	34	ND	ND	13.6
	35	ND	ND	17.0
	36	63.4	62.4	22.0
	37	56.3	55.8	22.5

35 \* HPLC conditions: Walters Delta Pak C-18 RP 2.5 $\mu$ m column; at 0 min 100% 0.1 TEAA; at 30 min 50% TEAA and 50% Acetonitrile; Flow rate 1.0 ml/min.

\*\* Tm at 4 $\mu$ M each strand from fit of duplicate melting curves to 2-state model with linear

sloping base line. Conditions: 100 mM NaCl, 10 mM Phosphate, 0.1 mM EDTA, pH 7.0.

ND = not determined

As is evident from Table 2, conjugates of cholic acid  
5 at the end of the oligonucleotide do not affect the Tm of the  
oligonucleotides.

2. Strands Incorporating 2'-O-Pentylamino Linker

a. Synthesis

An oligonucleotide of the sequence:

10 Oligomer 38: GCA' CCG GA'A' GGT A'CG A'G  
wherein A' represents a nucleotide functionalized to  
incorporate a pentylamine functionality at its 2'-position was  
synthesized in a one micromole scale utilizing the method of  
Example 7-B. The oligonucleotide was purified by reverse  
15 phase HPLC, detritylated and desaltsed on Sephadex G-25. PAGE  
gel analysis showed a single band. A further oligonucleotide,  
Oligomer 39, having the same sequence but without any 2'-O-  
amino linker was synthesis in a standard manner. A  
complementary DNA oligonucleotide of the sequence:

20 Oligomer 40: CCT GGC CTT CCA TGC TC.  
was also synthesized in a standard manner as was a  
complementary RNA oligonucleotide of the sequence:

Oligomer 41: CCU GGC CUU CCA UGC UC

b. Melting Analysis

25 Melting analysis was conducted as per the method of  
Procedure B-1-b. The results are shown in Table 4.

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TABLE 4

Melting Temperature Of The Hybridization Complex Of  
The Oligonucleotide And Its Complementary Strand

Oligomer	Tm <sup>*</sup>	
	DNA <sup>1</sup>	RNA <sup>2</sup>
38	54.5	58.0
39	60.6	56.9
<sup>*</sup> Tm at 4 μM each strand from fit of duplicate melting curves to 2-state model with linear sloping base line. Conditions: 100 mM NaCl, 10 mM Phosphate, 0.1 mM EDTA, pH 7.0.		
1 Against DNA complementary strand, Oligomer 40. 2 Against RNA complementary strand, Oligomer 41		

As is evident from Table 4 against the RNA complementary strand the change in Tm's between the strand having 2'-amino linkers thereon and the unmodified strand is 1.1 degrees (0.22 change per modification). Against the DNA strand, the change is -6.1 degrees (-1.2 change per modification). When compared to the parent unmodified oligonucleotide the 2'-amino linker-containing strand has a stabilizing effect upon hybridization with RNA and a destabilizing effect upon hybridization with DNA.

Compounds of the invention were tested for their ability to increase cellular uptake. This was determined by judging either their ability to inhibit the expression of bovine papilloma virus-1 (BPV-1) or an assay involving luciferase production (for HIV-1).

## PROCEDURE C

Determination of Cellular Uptake Judged By The Inhibition Of Expression of Bovine Papilloma Virus-1 (bpv-1) As Measured By an E2 Transactivation Assay

For this test, a phosphorothioate oligonucleotide analog of the sequence:

Oligomer 42: CTG TCT CCA TCC TCT TCA CT

was used as the basic sequence. This sequence is designed to be complementary to the translation initiation region of the E2 gene of bovine papilloma virus type 1 (BPV-1). Oligomer

42 served as the positive control and standard for the assay.

2122030

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Oligomer 3 (from Example 4 above) served as a second test compound. It has the same basic sequence except it is a phosphorothioate oligonucleotide and further it has a cholic acid moiety conjugated at the 3'-end of the oligonucleotide.

5 Oligomer 2 (from Example 2 above) served as a third test compound. Again it is of the same sequence, it is a phosphorothioate oligonucleotide and it has a cholic acid moiety conjugated at the 5'-end. Oligomer 5 (from Example 5 above) served as a fourth test compound. Once again it has 10 the same sequence, is a phosphorothioate oligonucleotide and it has a cholic acid moiety conjugated at both the 3'-end and 5'-end. A fifth test compound was a phosphorothioate oligonucleotide with no significant sequence homology with BPV-1. A sixth test compound was a further phosphorothioate 15 oligonucleotide with no significant sequence homology with BPV-1. The last test compound, the seventh test compound, was a phosphorothioate oligonucleotide with cholic acid conjugated to the 3'-end but having no significant sequence homology with BPV-1. Compounds five, six and seven served as negative 20 controls for the assay.

For each test I-38 cells were plated at  $5 \times 10^4$  cells per  $\text{cm}^2$  in 60 mm petri dishes. Eight hours after plating, medium was aspirated and replaced with medium containing the test oligonucleotide and incubated overnight. Following 25 incubation, medium was aspirated and replaced with fresh medium without oligonucleotide and incubated for one hour. Cells were then transfected by the CaPO<sub>4</sub> method with 2 ug of pE2RE-1-CAT. After a four hour incubation period cells were glycerol shocked (15% glycerol) for 1 minute followed by 30 washing 2 times with PBS. Medium was replaced with DMEM containing oligonucleotide at the original concentration. Cells were incubated for 48 hours and harvested. Cell lysates were analyzed for chloramphenicol acetyl transferase by standard procedures. Acetylated and nonacetylated <sup>35</sup>"C chloramphenicol were separated by thin layer chromatography and quantitated by liquid scintillation. The results are expressed as percent acetylation.

Two lots of the positive control compound were found to acetylate at a level of 29% and 30%. The negative controls, test compounds five, six and seven, were found to acetylate at 59%, 58% and 47%, respectively. The 3'-cholic acid conjugate test compound, Oligomer 3, was found to acetylate to 23%, the 5'-cholic acid conjugate test compound, Oligomer 2, was found to acetylate to 36% and the test compound conjugated at both the 3'-end and the 5'-end, Oligomer 5, was found to acetylate to 27%.

10 The results of this test suggests that placement of a cholic acid moiety at the 3'-terminus of an oligonucleotide increase the activity. This in turn suggests that the increased activity was the result of increased cellular membrane transport.

15 PROCEDURE D

Determination of Cellular Uptake Judged By Inhibition of pHIVluc With Cholic Acid Linked 2'-O-Methyl Substituted Oligonucleotides

For this test the absence of an oligonucleotide in a 20 test well served as the control. All oligonucleotides were tested as 2'-O-methyl analogs. For this test an oligonucleotide of the sequence:

Oligomer 43: CCC AGG CUC AGA  
wherein each of the nucleotides of the oligonucleotide includes 25 a 2'-O-methyl substituent group served as the basic test compound. The second test compound of the sequence:

Oligomer 44: 5'-CHA CCC AGG CUC AGA  
wherein CHA represents cholic acid and where each of the nucleotides of the oligonucleotide includes a 2'-O-methyl 30 substituent group, was also of the same sequence as the first test compound. This second test compound included cholic acid conjugated to its 5'-end and was prepared as per the method of Example 3 utilizing 2'-O-methyl phosphoramidite intermediates as identified in Example 7-C. The third test 35 compound of the sequence:

Oligomer 45: CCC AGG CUC AGA 3'-CHA

wherein CHA represents cholic acid and where each of the nucleotides of the oligonucleotide includes a 2'-O-methyl substituent group was also of the same sequence as the first test compound. The third test compound included cholic acid conjugated to its 3'-end and was prepared as per the method of Example 4 utilizing 2'-O-methyl phosphoramidite intermediates as identified in Example 7-C. The fourth test compound was a 2'-O-Me oligonucleotide of a second sequence:

Oligomer 46: GAG CUC CCA GGC

10 where each of the nucleotides of the oligonucleotide includes a 2'-O-methyl substituent group. The fifth test compound was of sequence:

Oligomer 47: 5'-CHA GAG CUC CCA GGC.

wherein CHA represents cholic acid and where each of the 15 nucleotides of the oligonucleotide includes a 2'-O-methyl substituent group. It was of the same sequence as the fifth test compound. This test compound included cholic acid conjugated to its 5'-end and was prepared as per the method of Example 3 utilizing 2'-O-methyl phosphoramidite 20 intermediates as identified in Example 7-C.

A sixth test compound was a randomized oligonucleotide of the sequence:

Oligomer 48: CAU GCU GCA GCC.

HeLa cells were seeded at  $4 \times 10^3$  cells per well in 6-25 well culture dishes. Test oligonucleotides were added to triplicate wells at 1  $\mu$ M and allowed to incubate at 37°C for 20 hours. Medium and oligonucleotide were then removed, cells washed with PBS and the cells were  $\text{CaPO}_4$  transfected. Briefly, 5  $\mu$ g of pHIVluc, a plasmid expressing the luciferase 30 cDNA under the transcriptional control of the HIV LTR constructed by ligating the KpnI/HindIII restriction fragments of the plasmids pT3/T7luc and pHIVpac (NAR 19(12)) containing the luciferase cDNA and the HIV LTR respectively, and 6  $\mu$ g of pcDEBtat, a plasmid expressing the HIV tat protein under the 35 control of the SV40 promoter, were added to 500  $\mu$ l of 250 mM  $\text{CaCl}_2$ , then 500  $\mu$ l of 2x HBS was added followed by vortexing. After 30 minutes, the  $\text{CaPO}_4$  precipitate was divided evenly

between the six wells of the plate, which was then incubated for 4 hours. The media and precipitate were then removed, the cells washed with PBS, and fresh oligonucleotide and media were added. Incubation was continued overnight. Luciferase activity was determined for each well the following morning. Media was removed, then the cells washed 2X with PBS. The cells were then lysed on the plate with 200  $\mu$ l of LB (1% Trit X-100, 25 mM Glycylglycine pH 7.8, 15 mM MgSO<sub>4</sub>, 4 mM EGTA, 1 mM DTT). A 75  $\mu$ l aliquot from each well was then added to a well 10 of a 96 well plate along with 75  $\mu$ l of assay buffer (25 mM Glycylglycine pH 7.8, 15 mM MgSO<sub>4</sub>, 4 mM EGTA, 15 mM KPO<sub>4</sub>, 1 mM DTT, 2.5 mM ATP). The plate was then read in a Dynatec multiwell luminometer that injected 75  $\mu$ l of Luciferin buffer (25 mM Glycylglycine pH 7.8, 15 mM MgSO<sub>4</sub>, 4 mM EGTA, 4 mM DTT, 1 mM luciferin) into each well, immediately reading the light emitted (light units).

The random sequence compound (Oligomer 48) and the other non-cholic acid-conjugated test compounds (Oligomers 43 and 46) had comparable activity. The 5'-conjugate of the 20 first sequence (Oligomer 44) also had activity comparable to the non-conjugated compounds. The 5'-conjugate of the second sequence (Oligomer 47) showed a three-fold increase in activity compared to the non-conjugated compounds and the 3'-conjugate of the first sequence (Oligomer 45) showed a further 25 3-fold increase in activity compared to Oligomer 47.

All the test cholic acid-bearing oligonucleotides showed significant inhibition of luciferase production compared to non-cholic acid-bearing oligonucleotides. This suggests that the increased activity was the result of 30 increased cellular membrane transport of the cholic acid-bearing test oligonucleotides.

**EXAMPLE 13**

**Retinoic Acid Conjugated Oligonucleotide**

**A. Retinoic Acid N-Hydroxysuccinimide Ester**

35 Anhydrous DMF (150 ml) was added to a mixture of retinoic acid (15 mmol, 4.5 g, Fluka) and N-hydroxysuccinimide

2122030

44

(5.25 g, 45 mmol). The mixture was stirred in the presence of argon. EDAC [ethyl-3-(3-dimethylamino)propyl carbodiimide] (4 ml, 25 nmol) was then added and this mixture was then stirred overnight. The solution was then evaporated to a yellow gum and dissolved in 200 ml ethylacetate and washed successively with 4% NaHCO<sub>3</sub> solution (200 ml) followed by saturated NaCl solution, dried over anhydrous MgSO<sub>4</sub>, and evaporated to yield the desired compound as a yellow solid in nearly 90% yield.

10       B. Retinol Phosphoramidite

All-<sup>14</sup>C-retinol (1 g) was vacuum dried and dissolved in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (10 ml) in an argon atmosphere. Diisopropylethylamine (2.65 ml, 21.5 mmol) was syringed in and the reaction mixture was cooled in an ice-bath. 2-cyanoethyl-N, N-diisopropylchlorophosphoramidate (2.5 g, 2.45 ml, 10.5 mmol) was slowly added by syringe under argon atmosphere. The reaction mixture was stirred for 30 min. at which time TLC (CH<sub>2</sub>Cl<sub>2</sub>:CH<sub>3</sub>OH:Et<sub>3</sub>N, 90:10:0.1) indicated complete conversion of the alcohol to its phosphoramidite. The reaction mixture was added to 100 ml of saturated NaHCO<sub>3</sub>, followed by washing the reaction flask with (2 x 25 ml) CH<sub>2</sub>Cl<sub>2</sub>. The CH<sub>2</sub>Cl<sub>2</sub> layer was separated and washed with 100 ml of saturated NaCl solution and dried over anhydrous MgSO<sub>4</sub> and evaporated into a yellow foam. The amidite was used directly in the DNA synthesizer without further purification since decomposition of this amidite was noted upon silica column purification.

C. Retinoic Acid Functionalized Oligonucleotide

Multiple batches of an oligonucleotide of the sequence: Oligomer 49:

30       T<sub>4</sub>G<sub>6</sub> G<sub>4</sub>A<sub>6</sub> C<sub>2</sub>C<sub>6</sub>G<sub>5</sub> T<sub>4</sub>A<sub>6</sub>G<sub>4</sub> C<sub>4</sub>G<sub>4</sub>A<sub>4</sub> G<sub>6</sub>G<sub>4</sub>C<sub>2</sub>-3'AL

wherein AL represents a 3'-aminolinker and "s" represents a phosphorothioate inter-nucleotide backbone linkage were synthesized as per the procedure of Example 4 on 10  $\mu$ mol scales in the standard manner on the DNA synthesizer utilizing phosphoramidite methodology employing 3'-amine-ON solid support available from Clontech. During the synthesis, the phosphorothioate backbone was formed by the Beaucage reagent.

2122030  
45

The oligonucleotide was deprotected and purified using standard protocols.

The 3'-aminolinker-oligonucleotide (Oligomer 49, 100 OD units, approximately 550 nmols) was dissolved in freshly prepared NaHCO<sub>3</sub> buffer (500 µl, 0.2M, pH 8.1) and treated with a solution of retinoic acid N-hydroxy succinimide ester (50 mg, 125 µmols) dissolved in 500 µl of DMF. The reaction mixture was covered with aluminum foil and left at 37°C bath overnight. It was then passed through a Sephadex G-25 column (1 x 40 cm) and the first eluant was collected, concentrated and passed again through another Sephadex G-25 column (1 x 40 cm) to remove excess Vitamin-A reagent. Concentration of the yellow oligonucleotide fractions followed by HPLC purification yielded the retinoic acid-oligonucleotide conjugate.

## 15 EXAMPLE 14

## Folic Acid Conjugated Oligonucleotide

A mixture of folic acid (30 mg, 68 µmols) and 1-hydroxybenzotriazole (30 mg, 222 µmols) was dissolved in 900 ml of dry DMF. To this solution, 50 ml of EDAC (312 µmols) was added. The resultant yellow viscous material was vortexed well and 500 ml from the solution was transferred into 100 O.D. units of the 3'-aminolinker oligonucleotide (Oligomer 49, 537 µmols) dissolved in 0.2M NaHCO<sub>3</sub> buffer. The yellow solution was vortexed, covered with aluminum foil and allowed to react for 16 hrs. The mixture was then loaded into a Sephadex G-25 column (1 x 40 cm). The oligonucleotide fraction was collected, concentrated and passed one more time through Sephadex G-25 column. The oligonucleotide fractions were concentrated and purified by reverse phase HPLC. The conjugate appeared as a broad peak centered around 32.5 min. while the oligonucleotide starting material had retention time of 30 min. (5% - 40% of 100% CH<sub>3</sub>CN over 60 min. as solvent B and 50 mM TEAA pH 7.0 as the solvent A in reverse phase HPLC). The conjugate was concentrated and passed 35 through Sephadex G-25 column again for desalting. Gel

2122030

46

analysis indicated a slower moving material than the starting oligonucleotide.

**EXAMPLE 15****Methyl Folate Conjugated Oligonucleotide**

5 In a like manner to Example 14, 5-methyl folate was also attached to Oligomer 49.

**EXAMPLE 16****Pyridoxal Conjugated Oligonucleotide**

10 The 3'-aminolinker-oligonucleotide (Oligomer 49, 20 O.D. units, approximately 110 nmols, based on the calculated extinction coefficient of  $1.828 \times 10^5$  at 260 nm) was dissolved in 100 microliters of water. 100  $\mu$ l of 1M NaOAc buffer (pH 5.0) was added followed by 5 mg of pyridoxal hydrochloride (24  $\mu$ mole) and 50  $\mu$ l of 60 mM NaCNBH<sub>3</sub> solution. The solution was 15 vortexed and left aside for overnight. It was then passed through a Sephadex G-25 column and further purified in an analytical HPLC column.

**EXAMPLE 17****Tocopherol Conjugated Oligonucleotide**

20 A. Vitamin E (Tocopherol)-hemisuccinate-NHS ester  $\alpha$ -Tocopherolhemisuccinate (Sigma, 5 g, 9.4 mmols) was treated with 3 equivalents of N-hydroxysuccinimide and 2 equivalents of EDAC as described under the vitamin-A NHS ester synthesis, Example 13-A above. Work up in the same manner as 25 Example 13 yielded the title compound as a light brown wax-like solid.

**B. Tocopherol Conjugated Oligonucleotide**

$\alpha$ -Tocopherol-hemisuccinate-NHS ester was treated with Oligomer 49 in the same manner described in Example 13-C for 30 the retinoic acid conjugation. The conjugate was obtained in nearly 50% yield.

**EXAMPLE 18****Synthesis of Uridine Based Aminolinkers****A. Preparation of 5'-dimethoxytrityl-2'-(O-pentyl-N-phthalimido)uridine phosphoramidite**

Utilizing the protocol of Wagner, et al., *J. Org. Chem.* 1974, 39, 24, uridine (45 g, 0.184 mol) was refluxed with di-n-butylinoxide (45 g, 0.181 mol) in 1.4 l of anhydrous methanol for 4 hrs. The solvent was filtered and the resultant 2',3'-O-dibutylstannylene-uridine was dried under vacuum at 100°C for 4 hrs to yield 81 g (93%).

The 2',3'-O-dibutyl stannylene-uridine was dried over  $P_2O_5$  under vacuum for 12 hrs. To a solution of this compound (20 g, 42.1 mmols) in 500 ml of anhydrous DMF were added 25 g (84.2 nmols) of N(5-bromopentyl)phthalimide (Trans World Chemicals, Rockville, Maryland) and 12.75 g (85 mmols) of cesium fluoride (CeF) and the mixture was stirred at room temperature for 72 hrs. The reaction mixture evaporated, coevaporated once with toluene and the white residue was partitioned between EtOAc and water (400 ml each). The EtOAc layer was concentrated and applied to a silica column (700 g). Elution with  $CH_2Cl_2$ - $CH_3OH$  (20:1 v/v) gave fractions containing a mixture of the 2'- and 3'- isomers of O-pentyl- $\omega$ -N-phthalimido uridine, in 50% yield.

The mixture was allowed to react with DMT chloride in dry pyridine at room temperature for 6 hrs.  $CH_3OH$  was used to quench excess DMT-Cl and the residue was partitioned between  $CH_2Cl_2$  containing 0.5%  $Et_3N$  and water. The organic layer was dried ( $MgSO_4$ ) and the residue was applied to a silica column. Elution with  $CH_2Cl_2$ : $CH_3OH$  (20:1, v/v) separated the 2' and 3' isomers.

The 2'-O-pentyl- $\omega$ -N-phthalimido-5'-DMT-uridine was converted to its phosphoramidite as per the procedure referenced in Example 7.

2122030

48

B. Preparation of 5'-dimethoxytrityl-2-(O-hexyl-N-phthalimido)uridine phosphoramidite

In a like manner to Example 18-A, using N-(6-bromohexyl) phthalimide, a 2'- six carbon aminolinker was introduced at the 2'-position of uridine.

C. Preparation of 5'-dimethoxytrityl-2-(O-decyl-N-phthalimido)uridine phosphoramidite

In a like manner to Example 18-A N-(10-bromodecyl)phthalimide was similarly used to introduce a 2'-ten carbon aminolinker in the nucleotide.

EXAMPLE 19

Synthesis of Cytidine Based Aminolinkers

A. Preparation of 5'-dimethoxytrityl-2-(O-propyl-N-phthalimido)cytidine phosphoramidite

The 5'-DMT protected 2'-O-functionalized cytidine phosphoramidite was prepared as per the procedure of Example 7 substituting cytidine for adenosine.

B. Preparation of oligonucleotides having a 2'-aminolinker bearing 3'-terminal nucleotide

The following oligonucleotides having phosphodiester inter-nucleotide linkages and a 2'-aminolinker at the 3' terminal nucleotide were synthesized:

Oligomer 50: GGG GUC UCC AGG GGA UCU GAC

Oligomer 51: TCT GAG TAG CAG AGG AGC TC

wherein C represents a nucleotide functionalized to incorporate a propyl-N-phthalimido functionality. Oligomer 50 is antisense to the Cap region of CMV and Oligomer 51 is antisense to an ICAM sequence. The oligonucleotides were synthesized on a 3  $\mu$ mol scale. Upon completion of synthesis they were deprotected using standard protocols and purified by reverse phase HPLC, detritylated and desalted.

**EXAMPLE 20**

Conversion Of An Oligonucleotide Having A 2'-Aminolinker To An Oligonucleotide Having A Thiolinker

**A. Oligomer 50**

5 Oligomer 50 (25 O.D. units) was treated with 5 mg SATA (N-succinimidyl-S-acetylthioacetate) in 0.2M NaHCO<sub>3</sub> buffer. The reaction mixture was passed through a Sephadex G-25 column, the oligonucleotide fraction was concentrated and treated with 200 mM NH<sub>2</sub>OH hydrochloride solution in water (1 10 ml).

**B. Oligomer 51**

Oligomer 51 (25 O.D. units) was treated with 5 mg SATA (N-succinimidyl-S-acetylthioacetate) in 0.2M NaHCO<sub>3</sub> buffer. The reaction mixture was passed through a Sephadex G-25 15 column, the oligonucleotide fraction was concentrated and treated with 200 mM hydroxylamine hydrochloride solution in water (1 ml).

**EXAMPLE 21**

Conjugation of o-Phenanthroline at 2'-Position Of 20 Oligonucleotides

**A. Oligomer 52**

To the solution resulting from Example 20-A was added 2 mg of 5-(iodoacetamide)-o-phenanthroline reagent followed by shaking overnight. The conjugate was purified by a size 25 exclusion column and reverse phase HPLC to yield

Oligomer 52: GGG GUC UCC AGG GGA UCU GAC-2'PHA  
wherein PHA represents a nucleotide functionalized at its 2'-position with phenanthroline via a thiol linker of the structure 2'-o-(CH<sub>2</sub>)<sub>3</sub>-NH-C(=O)-CH<sub>2</sub>-S-CH<sub>2</sub>-C(=O)-NH-.

**B. Oligomer 53**

To the solution resulting from Example 20-A was added 2 mg of 5-(iodoacetamide)-o-phenanthroline reagent followed by shaking overnight. The conjugate was purified by a size exclusion column and reverse phase HPLC to yield

35 Oligomer 53: TCT GAG TAG CAG AGG AGC TC-2'PHA

2122030

50

wherein PHA represents a nucleotide functionalized at its 2'-position with phenanthroline via a thiol linker of the structure 2'-O-(CH<sub>2</sub>)<sub>3</sub>-NH-C(=O)-CH<sub>2</sub>-S-CH<sub>2</sub>-C(=O)-NH-.

**EXAMPLE 22**

5 Oligonucleotide Having A Nucleotide With A Crosslinker/Aalkylator Attached Via A 2'-Aminolinker In A Internal Position In The Oligonucleotide

A. Synthesis of an oligonucleotide having a uridine 2'-aminolinker

10 The following oligonucleotide having phosphodiester inter-nucleotide linkages and a 2'-aminolinker at an internal position is synthesized utilizing the uridine 2'-aminolinker of Example 18:

Oligomer 54: GGC CAG AUC UGA GCC UGG GAG CU'C UGU GGC  
15 C

wherein U' represents a nucleotide functionalized to incorporate a propyl-N-phthalimido functionality. Oligomer 50 is an oligonucleotide corresponding to positions G<sub>16</sub> to C<sub>46</sub> of TAR RNA.

20 B. Conjugation of iodo acetamide to U<sub>38</sub> position of TAR structure

Oligomer 54 is reacted with iodoacetic acid N-hydroxy-succinimide ester to form the iodoacetamide derivative at the U<sub>38</sub> position of the TAR structure. The U<sub>38</sub> position is thus available for crosslinking to the 7 position of the guanine base of either G<sub>46</sub> or C<sub>28</sub> of the TAR structure.

**EXAMPLE 23**

Conjugation of Pyrene at 2'-Position Of Oligonucleotides

A. Single 2' site modification

30 10 O.D. units (A<sub>260</sub>) of Oligomer 12 (Example 7-B) (approximately 60 nmols based on the calculated extinction coefficient of 1.68 × 10<sup>5</sup>) was dried in a microfuge tube. It was dissolved in 200 µl of 0.2 M NaHCO<sub>3</sub> buffer and pyrene-1-butrylic acid N-hydroxysuccinimide ester (i.e., succinimidyl-1-

Pyrene butyrate, 3 mg, 7.79  $\mu$ moles, Molecular Probes, Eugene, Oregon) was added followed by 400  $\mu$ l of DMF. The mixture was incubated at 37°C overnight. The solution was applied to a Sephadex G-25 column (1 x 40 cm) and the oligonucleotide fractions were combined. The product was purified by HPLC. The pyrene conjugates exhibited the typical pyrene absorption between 300 and 400 nm. The product had a HPLC retention time of 26.94 min. while the parent oligonucleotide had a retention time of 21.78 min. (Waters 600E with 991 detector; Hamilton PRP-1 column (15x25 cm); Solvent A: 50 mM TEAA, pH 7.0; B: 45 mM TEAA with 80% Acetonitrile; 1.5 mL/min. flow rate; Gradient 5% B for first 5 minutes, linear (1%) increase in B every minute afterwards).

B. Multiple 2' site modifications

15 10 O.D. units of Oligomer 12 (Example 7-B) was treated with twice the amount of pyrene-1-butyrlic acid N-hydroxysuccinimide (6 mg in 400  $\mu$ l DMF) and worked up in the same fashion as Example 23-A. Sephadex G-25 purification followed by HPLC purification gave the doubly pyrene-  
20 conjugated oligonucleotide. The doubly conjugated oligonucleotide exhibited a HPLC retention time of 32.32 min. while the parent oligonucleotide had a retention time of 21.78 min. (Waters 600E with 991 detector; Hamilton PRP-1 column (15x25 cm); Solvent A: 50 mM TEAA, pH 7.0; B: 45 mM TEAA with  
25 80% Acetonitrile; 1.5 mL/min. flow rate; Gradient 5% B for first 5 minutes, linear (1%) increase in B every minute afterwards).

EXAMPLE 24

Conjugation of Acridine at 2'-Position Of Oligonucleotides

30 A. Single 2' site modification  
10 O.D. units ( $A_{260}$ ) of Oligomer 12 (Example 7-B, about 60 nmols) was dried and dissolved in 1M NaHCO<sub>3</sub>/Na<sub>2</sub>CO<sub>3</sub> buffer, pH 9.0, 200  $\mu$ l. 9-acridinyli-isothiocyanate, (5 mg, 2.1 nmols, Molecular Probes, Eugene, Oregon) was dissolved in 200  $\mu$ l DMF.  
35 This solution was added to the oligonucleotide, vortexed, covered with aluminum foil and left at 37°C overnight. The

reaction mixture was purified by passing through a Sephadex G-25 column (1 x 40 cm) concentrated and further purified by HPLC (reverse-phase). The product had a HPLC retention time of 25.32 min. while the parent oligonucleotide had a retention time of 21.78 min. (Waters 600E with 991 detector; Hamilton PRP-1 column (15x25 cm); Solvent A: 50 mM TEAA, pH 7.0; B: 45 mM TEAA with 80% Acetonitrile; 1.5 mL/min. flow rate; Gradient 5% B for first 5 minutes, linear (1%) increase in B every minute afterwards).

10       B. Multiple 2' site modifications

10 O.D. units ( $A_{260}$ ) of Oligomer 13 (Example 7-B) in 400  $\mu$ l of 1M Na<sub>2</sub>CO<sub>3</sub>/NaHCO<sub>3</sub> buffer (pH 9.0) was treated with 10 mg of 9-acridinyl-isothiocyanate in 400  $\mu$ l of DMF. The reaction mixture was vortexed, covered with aluminum foil and left at 15 37°C overnight. The reaction mixture was purified as for the single site reaction of Example 24-A. The doubly conjugated acridine-oligonucleotide eluted as the last peak in the HPLC following single-modification products. The product had a HPLC retention time of 32.32 min. while the parent oligo-20 nucleotide had a retention time of 21.78 min. (Waters 600E with 991 detector; Hamilton PRP-1 column (15x25 cm); Solvent A: 50 mM TEAA, pH 7.0; B: 45 mM TEAA with 80% Acetonitrile; 1.5 mL/min. flow rate; Gradient 5% B for first 5 minutes, linear (1%) increase in B every minute afterwards).

25 EXAMPLE 25

Conjugation Of Porphyrin at 2'-Position Of Oligonucleotides  
Methylpyrroporphyrin XXI ethyl ester (Aldrich) is condensed with aminocaproic acid using N-hydroxysuccinimide and EDAC. The resultant carboxylic acid is then activated 30 again with N-hydroxy succinimide and EDAC and treated with Oligomer 12 as per the procedure of Example 23-A to give the 2'-porphyrin conjugated oligonucleotide.

**EXAMPLE 26****Conjugation of Hybrid Intercalator-Ligand at 2'-Position of Oligonucleotides****A. Photonuclease/intercalator ligand**

The photonuclease/intercalator ligand 6-[[(9-[(6-(4-nitrobenzamido)hexyl]amino)acridin-4-yl]carbonyl]amino]hexanoylpentafluorophenyl ester was synthesized as per the procedure of Egholm et al., *J. Am. Chem. Soc.* 1992, 114, 1895.

**B. Single 2' site modification**

10 O.D. units of Oligomer 12 (Example 7-B) was dissolved in 100  $\mu$ l of 0.1 M borate buffer (pH 8.4) and treated with 330  $\mu$ l of DMF solution (10 mg in 1 ml of DMF) of 6-[[(9-[(6-(4-nitrobenzamido)hexyl]amino)acridin-4-yl]carbonyl]amino]hexanoylpentafluorophenyl ester. The solution was covered with aluminum foil and allowed to react overnight. The product was purified by Sephadex G-25 and HPLC purification of the reaction mixture.

**C. Multiple 2' site modification**

20 10 O.D. units  $A_{260}$  of Oligomer 13 (Example 7-B) was dissolved in 200  $\mu$ l of 0.1 M borate buffer (pH 8.4) and treated with 660  $\mu$ l of the DMF solution of 6-[[(9-[(6-(4-nitrobenzamido)hexyl]amino)acridin-4-yl]carbonyl]amino]hexanoylpentafluorophenyl ester (10 mg in 25 1 ml solution) and the solution was covered with aluminum foil and left aside overnight. The bright yellow solution was purified by Sephadex G-25 and reverse phase HPLC to give the doubly conjugated oligonucleotide.

**EXAMPLE 27****Conjugation of Bipyridine complex at 2'-Position of Oligonucleotides****A. Bipyridine complex**

Succinimidyl-4'-carboxyl-2,2'-bipyridine is synthesized according to the procedure of Telser, et al., *J. Am. Chem. Soc.* 1989, 111, 7221.

2122030

54

## B. Single 2' site Modification

10 O.D. A<sub>260</sub> units of Oligomer 12 is reacted with a 200 fold molar excess of succinimidyl-4-carboxyl-4'-methyl-2',2'-bipyridine in 0.1 M borate buffer pH 8.5/DMF. The solution is purified by Sephadex G-25 and reverse phase HPLC to yield the conjugated oligonucleotide.

## EXAMPLE 28

## Conjugation of Aryl Amide Photocrosslinkers at 2'-Position Of Oligonucleotides

## 10 A. Conjugation of N-hydroxysuccinimidyl-4-azidobenzoate (HSAB)

Oligomer 14 (i.e., TTC CTT CCA\* TCT TCC TCG TC wherein A\* represents 2'-O-pentyl amino adenosine, Example 7-C, 100 O.D. units, 595 nmols, based on the calculated extinction coefficient of  $1.6792 \times 10^6$  OD units) was dried and dissolved in 500 ml of 0.2M NaHCO<sub>3</sub> buffer pH 8.1 and treated with 25 mg of N-hydroxysuccinimidyl-4-azidobenzoate (HSAB, 96  $\mu$ moles, available both from Pierce & Sigma) dissolved in 500  $\mu$ l of DMF. The reaction was allowed to react overnight at 37°C and 20 passed twice over Sephadex G-25 column (1 x 40 cm). The oligonucleotide fraction was purified by reverse-phase HPLC. The product had the HPLC retention time of 38.79 min while the parent oligonucleotide had the retention time of 33.69 min. ( $\pm 40\%$  CH<sub>3</sub>CN in 60 min.) in reverse phase column.

## 25 B. Conjugation of N-succinimidyl-6(4'-azido-2'-nitrophenyl-amino)hexanoate

Oligomer 14 (Example 7-C, 200 OD units) was dissolved in 500 ml NaHCO<sub>3</sub> buffer (0.2M, pH 8.1) and treated with 500 mg of N-succinimidyl-6(4'-azido-2'-nitrophenyl-amino)hexanoate (SANPAH, 128  $\mu$ mole, available both from Pierce and Sigma) dissolved in 500  $\mu$ l DMF. The reaction vial was wrapped with aluminum foil and heated at 37°C overnight. The reaction mixture was passed twice over a Sephadex G-25 column (1 x 40 cm). The oligonucleotide fraction was purified by reverse-phase HPLC. The product had the HPLC retention time of 40.69 min. while the parent oligonucleotide had the retention time

of 33.69 min. (5% → 40% CH<sub>3</sub>CN in 60 min.) in a reverse phase column.

PROCEDURE D

Duplex Melting Temperature of Conjugated Oligonucleotides

Utilizing the protocol described in Procedure B-1-b, the melting temperatures of various of the 2'-aminolinked conjugate oligomers of the invention against a complementary DNA strand were obtained. The conjugate oligomers were compared to an oligomer of the same sequence bearing a 2'-O-pentylamino group. An un-modified, i.e., wild type, strand of the same sequence was also tested for comparison purposes. Both single site and multiple site conjugated oligomers were tested. As is shown in Table 5, the T<sub>m</sub> and the A/T<sub>m</sub>/modification, both as compared to 2'-pentylamino bearing oligomer, were measured. The wild type sequence is:

Oligomer 55:

CTG TCT CCA TCC TCT TCA CT  
the single site sequence is:

Oligomer 12:

CTG TCT CCA<sup>\*</sup> TCC TCT TCA CT  
and the multiple site sequence is:

Oligomer 13:

CTG TCT CCA<sup>\*</sup> TCC TCT TCA<sup>\*</sup> CT  
where A<sup>\*</sup> represents a site of conjugation.

56

TABLE 5

Duplex Melting Temperature of  
Conjugated Oligonucleotides Against DNA

Oligomer	Conjugate	T <sub>m</sub> °C	ΔT <sub>m</sub> /mod
5	wild type	60.5	--
12	2'-O-pentyl-NH <sub>2</sub>	58.1	--
12	biotin	56.4	-1.7
12	cholic acid	55.5	-2.6
10	digoxigenin	55.8	-2.3
12	fluorescein	55.1	-3.0
12	pyrene	62.6	+4.5
12	acridine	58.6	+0.5
13	2'-O-pentyl-NH <sub>2</sub>	56.9	--
15	biotin	54.4	-1.3
13	cholic acid	54.3	-1.3
13	digoxigenin	53.8	-1.6
13	fluorescein	53.4	-1.8
13	pyrene	65.1	+4.1
20	acridine	58.1	+1.2

## EXAMPLE 29

Conjugation of Imidazole-4-acetic acid at 2'-Position of  
Oligonucleotides

## A. Activated imidazole-4-acetic acid

25 Imidazole-4-acetic acid was reacted with 2,4-dinitrofluoro benzene. The resulting imidazole-N-(DNP)-4-acetic acid was converted to its N-hydroxy succinimide ester by treating with NHS/EDAC as per the procedure of Example 13.

## B. 2'-Site Modification

30 10 O.D. A<sub>260</sub> units of oligomer 12 was reacted with a 100 fold molar excess of imidazole-N-(DNP)-4-acetic acid NHS ester in 0.1M borate buffer pH 8.5/DMF (200 μL each). After overnight reaction, the reaction mixture was treated with 200 μL of mercaptoethanol to cleave off the DNP protecting group.

35 The resulting reaction mixture was purified by passing through a Sephadex G-25 column followed by reverse phase HPLC to yield the imidazole conjugated oligonucleotide.

2122030

**EXAMPLE 30****Conjugation of Metal Chelating Agents to 2'-Position of Oligonucleotide****A. EDTA Complex**

5 To form an EDTA Fe(II) complex for coupling to an oligonucleotide as a nucleic acid cleaving agent, tricyclohexyl ester of EDTA is synthesized according to the procedure of Sluka, et al., J. Am. Chem. Soc. 1990, 112, 6369.

**B. EDTA 2'-Site Modification**

10 The tricyclohexyl ester of EDTA (1.25 mg, 1.94 mmol) and hydroxybenzotriazole (HOBT, 1 mg, 6.6 mmol) are dissolved in DMF (50  $\mu$ L) and EDAC 10  $\mu$ L is added. To this solution, oligonucleotide 12 (10 OD units) in 100  $\mu$ L g 0.1M borate buffer is added and left overnight. The solution is passed 15 through a Sephadex G-25 column and the oligonucleotide fraction treated with conc. NH<sub>3</sub> (100  $\mu$ L) for 1 hr. to cleave off the acid protecting groups. Finally purification is effected by size exclusion and HPLC.

**C. DTPA 2'-Site Modification**

20 Oligomer 12 was treated with diethylenetriamine pentaacetic anhydride (DTPA) in 0.1M NaHCO<sub>3</sub>/DMF to offer single-site modification. The conjugate was complexed with Gadolinium ion (Gd III) to give a contrast agent, usable among other uses, as an uptake measuring agent.

**25 EXAMPLE 31****Conjugation of Cholesterol To The 2'-Position of Oligonucleotide****A. Method 1 - 2'-Aminolinker**

Cholesterol-hemisuccinate was converted to its N-30 hydroxy succinimide ester. It was then conjugated to Oligomer 12 as per the procedure of Example 23-A or Oligomer 13 as per the procedure of Example 23-B.

2122030

58

B. Method 2 - Conjugation of Cholesterol via a Disulfide Bridge

Step 1

Thiocholesterol (1.4 g, 3.5 mmol) is added to a stirred solution of 2,2'-dithiobis(5-nitropyridine) (1.4 g 4 mmol) in chloroform 20 mL containing glacial acetic acid (400  $\mu$ L) under an argon atmosphere. The reaction is allowed to continue overnight at room temperature, after which the precipitated 5-nitropyridine-2-thione was removed and the solution 10 evaporated and purified in a silica column to give S-(2-thio-5-nitropyridyl)-thio cholesterol.

Step 2

Oligomer 55:

T<sub>n</sub>G<sub>n</sub>, G<sub>n</sub>A<sub>n</sub>, C<sub>n</sub>C<sub>n</sub>, T<sub>n</sub>A<sup>n</sup>G<sub>n</sub>, C<sub>n</sub>G<sub>n</sub>A<sub>n</sub>, G<sub>n</sub>C<sub>n</sub>,

15 wherein A represents an adenosine nucleotide functionalized to incorporate a 2'-O-pentylamino linking group is synthesized as per Example 7. This oligonucleotide is then converted into a thiol linker compound as per the procedure described for Oligomer 50 in Example 20.

20 The thiol linker group containing oligonucleotide, Oligomer 55, is reacted with an excess of S-(2-thio-5-nitropyridyl)-thiocholesterol to conjugate the cholesterol moiety to the 2' position of the oligonucleotide via a disulfide bridge.

25 EXAMPLE 32

Synthesis of 2'-Aminolinker Containing Solid Supports for DNA/RNA Synthesis

A. 5'-Dimethoxytrityl-2'-O-(pentyl-N-phthalimido)uridine

30 5'-Dimethoxytrityl-2'-O-(pentyl-N-phthalimido)uridine was synthesized as per the procedure described in Example 18.

B. Succinate Nucleoside

The nucleoside of step A (1 mmol) was treated with 4-DMAP (122 mg, 1 mmol) and succinic anhydride (250 mg, 2.5 mmol) in 10 mL g anhydrous pyridine. After shaking overnight, 35 TLC (EtOAc: Hexane 6:4 with 0.1% Et<sub>3</sub>N) indicated complete

2122030  
59

succinylation of the nucleoside. 10 mL of water was added and the reaction shaken for an additional 1 hr. The reaction mixture was evaporated and partitioned between CHCl<sub>3</sub> and 20% citric acid (50 mL each). The chloroform layer was washed 5 with brine and evaporated. It was then used in the next step.

C. Nitrophenyl Succinate

The dry 3'-O-succinate from step B was dissolved in dry dioxan (10 mL) containing pyridine (400 mL). 4-Nitrophenol (280 mg, 2 mmol) was added followed by DCC (1.32 g, 5 mmol) and the solution was shaken for 24 hrs. The fine precipitate of urea was filtered and the filtrate evaporated and applied to a silica column. The column was eluted with 5% CH<sub>2</sub>OEt in CHCl<sub>3</sub> containing 0.1% Et<sub>3</sub>N. The 3'-nitrophenyl succinate of the nucleoside eluted first from the column. The 15 product containing fractions were evaporated to give a foam.

D. Nucleoside Solid Support

The 3'-nitrophenyl succinate from step C was dissolved in 5 mL of anhydrous DMF and treated with 5g of 500 Å pore diameter aminopropyl CPG support and shaken for 8 hrs. The 20 CPG support was filtered, washed with methanol (5 x 20 mL) followed by ether (5 x 20) and dried. The CPG support was capped by treating for 30 min. with pyridine/acetic anhydride/N-methyl imidazole (20 mL, 8:1:1 v/v/v). The CPG support was then filtered off, washed with pyridine, methanol 25 and ether and air-dried. Assay of the dimethoxy trityl showed the loading capacity of the CPG support was 27 mmols/gram.

PROCEDURE E

Determination of Cellular Uptake of Folic Acid Conjugated Oligonucleotide

30 The effect of conjugation of an oligonucleotide with folic acid was determined by the inhibition of ICAM-1 utilizing the method of Chiang, et al., J. Biol. Chem. 1991, 266, 18162. Utilizing this method, human lung epithelial carcinoma cells (A549 cells) were grown to confluence in 96 35 well plates. Medium was removed and the cells were washed with folic acid free medium three times. Folic acid free

medium was added to the cells and increasing concentrations of an ICAM-1 specific antisense phosphorothioate oligonucleotide having the sequence 5'-TGG GAG CCA TAG CGA GGC-3', either free or conjugated to folic acid, was added to 5 the incubation medium. This oligonucleotide is an 18 base phosphorothioate oligonucleotide that targets the AUG translation initiation codon of the human ICAM-1 mRNA (Chiang et al., J. Biol. Chem. 1991, 266, 18162). The oligonucleotides were incubated with the A549 cells for 24 10 hours then ICAM-1 was induced by adding 2.5 ng/ml tumor necrosis factor- $\alpha$  to the medium. Cells were incubated an additional 15 hours in the presence of tumor necrosis factor- $\alpha$  and oligonucleotide. ICAM-1 expression was determined by a specific ELISA as described by Chiang, et al.. We had 15 previously demonstrated that the addition of the test oligonucleotide to incubation medium alone does not result in inhibition of ICAM-1 expression. However formulation of the test oligonucleotide with cationic liposomes results in at least a 1000 fold increase in potency and also correlates with 20 the appearance of the oligonucleotide in the nucleus (Bennett, et al., Molecular Pharmacology 1991, 41, 1023). The results of this test are shown in Table 6. At the 30uM level, the folic acid conjugated oligonucleotide shows an approximate 40% enhancement in activity.

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TABLE 6

## ICAM-1 Activity

Concentration	Oligonucleotide Control	Oligonucleotide + Folic Acid - Percent Of Control
1 uM	105.1	107.6
3 uM	112.0	104.4
10 uM	112.4	92.9
30 uM	108.4	61.6

93/07883

2122030 PCT/US92/09196

61

Those skilled in the art will appreciate that numerous changes and modifications may be made to the preferred embodiments of the invention and that such changes and modifications may be made without departing from the spirit 5 of the invention. It is therefore intended that the appended claims cover all such equivalent variations as fall within the true spirit and scope of the invention.

2122030

62

## WE CLAIM:

1. A compound comprising a plurality of linked nucleosides, wherein at least one of said nucleosides is functionalized at its 2'-position by attachment of a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, a lipid soluble vitamin, an RNA cleaving complex, a metal chelator, a porphyrin, an alkylator, a hybrid photonuclease/intercalator, or an aryl azide photo-crosslinking agent.
2. The compound of claim 1 wherein a linking moiety attaches said steroid molecule, reporter molecule, non-aromatic lipophilic molecule, reporter enzyme, peptide or protein to the 2'-position of said functionalized nucleoside.
3. The compound of claim 1 wherein said functionalized nucleoside comprises a steroid molecule, biotin, a fluorescein dye molecule, an alicyclic hydrocarbon molecule, a saturated or unsaturated fatty acid, a wax, a terpene, a polycyclic hydrocarbon, adamantane, a buckminsterfullerene, a reporter enzyme, a peptide, a protein, thiamine, riboflavin, nicotinic acid, niacin, pyridoxal, pantothenic acid, biotin, folic acid, methyl folate, a vitamin B<sub>12</sub> cobamide coenzyme, inositol, choline, ascorbic acid, retinoic acid, retinol, a tocopherol, vitamin D, vitamin K, EDTA, DTPA, o-phenanthroline, a photonuclease/intercalator, an aryl azide photo-crosslinker, a phenanthroline/metal complex and a bipyridine/metal complex attached to the 2'-position of said nucleoside.
4. The compound of claim 1 wherein said functionalized nucleoside comprises a steroid molecule attached via a linking moiety to the 2'-position of said nucleoside.

2122030

63

5. The compound of claim 2 wherein said linking moiety comprises an N-aminoalkoxy or N-aminoalkylamino moiety.

6. The compound of claim 5 wherein said linking moiety comprises an N-aminoalkoxy moiety.

7. The compound of claim 1 wherein said functionalized nucleoside comprises a reporter enzyme, a peptide or protein attached via a linking moiety to the 2'-position of said nucleoside.

8. The compound of claim 1 wherein cholic acid, dehydrocholic acid, cortisone, digoxigenin, testosterone, cholesterol or digoxigenin is attached at the 2' position of said nucleoside.

9. The compound of claim 8 wherein attachment is through a linking moiety comprises an N-aminoalkoxy or N-aminoalkylamino moiety.

10. The compound of claim 9 wherein said linking moiety comprises an N-aminoalkoxy moiety.

11. The compound of claim 10 wherein said linking moiety comprises a 5-aminopentoxo group.

12. The compound of claim 1 wherein cholic acid is attached at the 2' position of said nucleoside.

13. The compound of claim 12 wherein said attachment is through a linking moiety comprises an N-aminoalkoxy or N-aminoalkylamino moiety.

14. The compound of claim 13 wherein said linking moiety comprises an N-aminoalkoxy moiety.

2122030 64

15. The compound of claim 14 wherein said linking moiety comprises a 5-aminopentoxy group.

16. The compound of claim 1 wherein at least one of the linked nucleosides is a 2'-deoxy-2'-fluoro, 2'-methoxy, 2'-ethoxy, 2'-propxy, 2'-aminoalkoxy or 2'-allyloxy nucleoside.

17. The compound of claim 1 wherein at least two of the linked nucleosides are linked by phosphorothioate linking groups.

18. A compound comprising a plurality of linked nucleosides, wherein at least one of the nucleosides is a functionalized nucleoside selected from the group consisting of:

a 2' functionalized nucleoside having cholic acid linked to the 2' position of the nucleoside;  
a heterocyclic base functionalized nucleoside having cholic acid linked to the heterocyclic base of the nucleoside;  
a 5' terminal nucleoside having cholic acid linked to the 5'-position of the nucleoside;  
a 3' terminal nucleoside having cholic acid linked to the 3'-position of the nucleoside; and  
an inter-strand nucleoside having cholic acid linked to an inter-nucleotide linkage linking said inter-strand nucleoside to an adjacent nucleoside.

19. The compound of claim 18 wherein a linking group links said cholic acid and the functionalized nucleoside.

20. The compound of claim 18 wherein said functionalized nucleoside is a 2' functionalized nucleoside having cholic acid linked to the 2'-position of the nucleoside.

2122030  
65

21. The compound of claim 18 wherein at least one of the linked nucleosides is a 2'-deoxy-2'-fluoro, 2'-methoxy, 2'-ethoxy, 2'-propoxy, 2'-aminoalkoxy or 2'-allyloxy nucleoside.

22. The compound of claim 18 wherein at least two of the linked nucleosides are linked by phosphorothioate linking groups.

23. The compound of claim 18 wherein said functionalized nucleoside is a 3' terminal nucleoside having cholic acid linked to the 3'-position of said 3' terminal nucleoside.

24. The compound of claim 18 wherein said functionalized nucleoside is a 5' terminal nucleoside having cholic acid linked to the 5'-position of said 5' terminal nucleoside.

25. The compound of claim 18 wherein said functionalized nucleoside is a heterocyclic base functionalized nucleoside having cholic acid linked to the heterocyclic base of said nucleoside.

26. The compound of claim 18 wherein said functionalized nucleoside is an inter-strand nucleoside having cholic acid linked to an inter-nucleotide linkage linking said inter-strand nucleoside to an adjacent nucleoside.

27. A compound comprising a plurality of linked nucleosides, wherein:

at least one of the nucleosides is a 2'-deoxy-2'-fluoro, 2'-O-C<sub>1</sub>-C<sub>20</sub>-alkyl, 2'-O-C<sub>2</sub>-C<sub>20</sub>-alkenyl, 2'-O-C<sub>2</sub>-C<sub>20</sub>-alkynyl, 2'-S-C<sub>1</sub>-C<sub>20</sub>-alkyl, 2'-S-C<sub>2</sub>-C<sub>20</sub>-alkenyl, 2'-S-C<sub>2</sub>-C<sub>20</sub>-alkynyl, 2'-NH-C<sub>1</sub>-C<sub>20</sub>-alkyl, 2'-NH-C<sub>2</sub>-C<sub>20</sub>-alkenyl, 2'-NH-C<sub>2</sub>-C<sub>20</sub>-alkynyl nucleoside; and

2122030

66

at least one of the nucleosides is a functionalized nucleoside selected from the group comprising::

a 2'-functionalized nucleoside having a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, a lipid soluble vitamin, an RNA cleaving complex, a metal chelator, a porphyrin, an alkylator, a hybrid photonuclease/intercalator, or an aryl azide photo-crosslinking agent linked to the 2'-position of the nucleoside;

a heterocyclic base functionalized nucleoside having a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, a lipid soluble vitamin, an RNA cleaving complex, a metal chelator, a porphyrin, an alkylator, a hybrid photonuclease/intercalator, or an aryl azide photo-crosslinking agent linked to the heterocyclic base of the nucleoside;

a 5' terminal nucleoside having a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, a lipid soluble vitamin, an RNA cleaving complex, a metal chelator, a porphyrin, an alkylator, a hybrid photonuclease/intercalator, or an aryl azide photo-crosslinking agent linked to the 5'-position of the nucleoside;

a 3' terminal nucleoside having a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, a lipid soluble vitamin, an RNA cleaving complex, a metal chelator, a porphyrin, an alkylator, a hybrid photonuclease/intercalator, or an aryl azide photo-crosslinking agent linked to the 3'-position of the nucleoside; and

an inter-strand nucleoside having a steroid molecule, a reporter molecule, a non-aromatic

2122030

lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, a lipid soluble vitamin, an RNA cleaving complex, a metal chelator, a porphyrin, an alkylator, a hybrid photonuclease/intercalator, or an aryl azide photo-crosslinking agent linked to an inter-strand linkage linking said inter-strand nucleoside to an adjacent nucleoside.

28. The compound of claim 27 wherein:

said functionalized nucleoside is a nucleoside having a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, a lipid soluble vitamin, an RNA cleaving complex, a metal chelator, a porphyrin, an alkylator, a hybrid photonuclease/intercalator, or an aryl azide photo-crosslinking agent linked to the 2'-position of said nucleoside; and

the remainder of said linked nucleosides are 2'-deoxy-2'-fluoro, 2'-methoxy, 2'-ethoxy, 2'-propoxy, 2'-aminoalkoxy or 2'-allyloxy nucleosides.

29. The compound of claim 27 wherein said functionalized nucleoside is a 3' terminal nucleoside having a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, a lipid soluble vitamin, an RNA cleaving complex, a metal chelator, a porphyrin, an alkylator, a hybrid photonuclease/intercalator, or an aryl azide photo-crosslinking agent linked to the 3'-position of the nucleoside.

30. The compound of claim 27 wherein said functionalized nucleoside is a 5' nucleoside having a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, a lipid soluble vitamin, an RNA cleaving complex, a metal chelator, a porphyrin, an alkylator, a hybrid

2122030

68

photonuclease/intercalator, or an aryl azide photo-crosslinking agent linked to the 5'-position of the nucleoside.

31. The compound of claim 27 wherein said functionalized nucleoside is a heterocyclic base functionalized nucleoside having a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, a lipid soluble vitamin, an RNA cleaving complex, a metal chelator, a porphyrin, an alkylator, a hybrid photonuclease/intercalator, or an aryl azide photo-crosslinking agent linked to the heterocyclic base of the nucleoside.

32. The compound of claim 27 wherein said functionalized nucleoside is an inter-strand nucleoside having a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, a lipid soluble vitamin, an RNA cleaving complex, a metal chelator, a porphyrin, an alkylator, a hybrid photonuclease/intercalator, or an aryl azide photo-crosslinking agent linked to an inter-strand linkage linking said nucleoside to an adjacent nucleoside.

33. The compound of claim 27 wherein at least two of the linked nucleosides are linked by phosphorothioate linking groups.

63189-337

2122030

69

34. Use of a compound having a plurality of linked nucleosides, wherein said compound includes at least one nucleoside functionalized at its 2'-position by attachment of a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, or a lipid soluble vitamin to the 2'-position with a linking moiety to effect cellular uptake of said compound in an organism.

35. The use of claim 34 further including selecting said steroid molecule from the group consisting of cholic acid, deoxycholic acid, dehydrocholic acid, cortisone, digoxigenin, testosterone, cholesterol and digoxigenin.

36. The use of claim 34 further including selecting cholic acid as said steroid molecule.

37. The use of claim 34 further including selecting an  $\alpha$ -aminoalkylamino moiety as said linking molecule.

38. A method of derivatizing an oligonucleotide comprising:  
functionalizing said oligonucleotide with at least one of a steroid molecule, a reporter molecule, a non-aromatic lipophilic molecule, a reporter enzyme, a peptide, a protein, a water soluble vitamin, a lipid soluble vitamin, an RNA cleaving complex, a metal chelator, a porphyrin, an alkylator,

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63189-337

2122030

69a

a hybrid photonuclease/intercalator, or an aryl azide photo-crosslinking agent; and

further functionalizing said oligonucleotide with one of a group to effect increased affinity of said oligonucleotide for a target or a group to increase the stability of said oligonucleotide to nucleases.

39. The method of claim 38 wherein said oligonucleotide is functionalized with one of a steroid molecule, biotin, a fluorescein dye molecule, an alicyclic hydrocarbon molecule, a saturated or unsaturated fatty acid, a wax, a terpene, a polycyclic hydrocarbon, adamantane, a buckminsterfullerene, a reporter enzyme, a peptide, a protein, pyrene, thiamine, riboflavin, nicotinic acid, niacin, pyridoxal pantothenic acid, biotin, folic acid, methyl folate, a vitamin b<sub>12</sub> cobamide coenzyme, inositol, choline, ascorbic acid, retinoic acid, retinol, a tocopherol, vitamin d, vitamin K, EDTA, DTPA, o-phenanthroline, a photonuclease/intercalator, a bipyridine complex, an aryl

2122030

70

azide photo-crosslinker, a phenanthroline/metal complex or a bipyridine/metal complex.

40. The method of claim 38 including further functionalizing said oligonucleotide to both increase affinity of said oligonucleotide for a target and to increase the stability of said oligonucleotide to nucleases.

41. A compound comprising a plurality of linked nucleosides, wherein at least one of the nucleosides is a functionalized nucleoside selected from the group consisting of:

- a 2' functionalized nucleoside having folic acid linked to the 2' position of the nucleoside;
- a heterocyclic base functionalized nucleoside having folic acid linked to the heterocyclic base of the nucleoside;
- a 5' terminal nucleoside having folic acid linked to the 5'-position of the nucleoside;
- a 3' terminal nucleoside having folic acid linked to the 3'-position of the nucleoside; and
- an inter-strand nucleoside having folic acid linked to an inter-nucleotide linkage linking said inter-strand nucleoside to an adjacent nucleoside.

42. A compound of claim 41 wherein said functionalized nucleoside is a 2' functionalized nucleoside having folic acid linked to the 2' position of the nucleoside.

43. A compound comprising a plurality of linked nucleosides, wherein at least one of the nucleosides is a functionalized nucleoside selected from the group consisting of:

- a 2' functionalized nucleoside having pyrene linked to the 2' position of the nucleoside;
- a 5' terminal nucleoside having pyrene linked to the 5'-position of the nucleoside;

a 3' terminal nucleoside having pyrene linked to the 3'-position of the nucleoside; and  
an inter-strand nucleoside having pyrene linked to an inter-nucleotide linkage linking said inter-strand nucleoside to an adjacent nucleoside.

44. A compound of claim 43 wherein said functionalized nucleoside is a 2' functionalized nucleoside having pyrene linked to the 2' position of the nucleoside.

